



Biosurfactants from renewable sources - A review

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(Received: 29 June 2021; Revised: 11 November 2022; Accepted: 30 December 2022)

Abstract

Biosurfactants have wide applications in pharmaceutical, agriculture and food industries. The research area of biosurfactants is gaining immense attention. The review mentions here the advantages and various substrates used for biosurfactants production. The pre-treatment of substrates for biosurfactants production is also focused. The production of biosurfactants by solid state fermentation is also described. The renewable substrates, yield and microorganisms used for biosurfactant production are also taken into consideration. The screening methods for biosurfactant are also described. The use of renewable sources for biosurfactant production is specially focused in the review. This will be very eco-friendly, easy and economical. More studies need to be done on large-scale production of biosurfactants using genetically engineered microorganisms.

Keywords: Agroindustry by-products, economical, eco-friendly, solid-state fermentation, value-added products

Biosurfactants

Biosurfactants are amphiphilic compounds which can be produced from microbes using cheap renewable substrates. The research on biosurfactants is gaining wide attention by the scientists as because biosurfactants are eco-friendly and economical and can be produced easily using raw substrates or wastes. Biosurfactants have immense applications in medical, pharmaceutical, food, dairy, etc. industries (Akbari et al., 2018). Much of the wastes or by-products are generated from industries such as food, dairy, agricultural, etc. These wastes are disposed to the landfills without any treatment. So, such by-products

or wastes can be used to produce biosurfactants. These wastes or by-products include bagasse, press mud, vegetable, fruit wastes such as orange, apple, banana peels, etc. These wastes or by-products generated in huge amounts are lignocellulosic in nature.

Types of biosurfactants (Roy, 2017)

The different biosurfactants include glycolipids, rhamnolipids, sophorolipids, trehalolipids, surfactin, lipopeptides and lipoproteins, polymeric and particulate (Fig. 1).

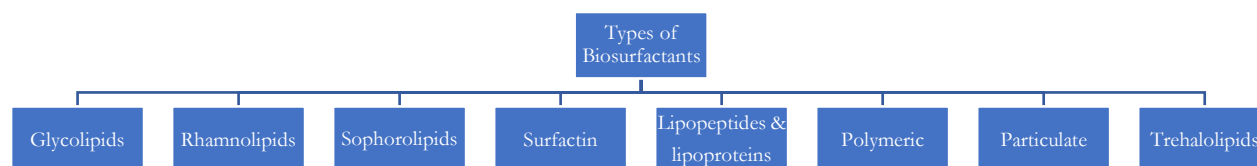


Figure 1 Types of biosurfactants

Advantages of biosurfactants (Fenibo et al., 2019)

- Easy to breakdown
- Non-toxic
- Biosurfactants are compatible and hence have wide applications in various industries.
- They can be produced using economical substrates or wastes.
- They can minimize the surface tension
- Biosurfactants have stability over a wide range of temperature, sodium chloride concentration and pH.
- They have specificity.

Global biosurfactant market

Biosurfactants have immense demand in the market. Surfactants have immense demand globally. The global market in 2016 for biosurfactant was USD 30.64 billion which raised to USD 39.86 billion by 2021 (Markets & Markets, 2016). The CAGR for biosurfactant in the market was 4.3% in 2020 (Grand View Research, 2015). By 2023, the biosurfactant market is said to reach USD 2.6 billion (Global Market Insights, 2018). Europe is the biggest market for biosurfactant production to grow as the biggest market (around 53%) and then USA. Sophorolipids, type of biosurfactants has the biggest market worldwide. Even

the synthetic biosurfactants are cheap with the market price \$2 kg⁻¹ globally (Santos et al., 2016a).

Substrates for biosurfactant production

The substrates used for biosurfactant production are agricultural, industrial, starch-rich, vegetable oil processing wastes, and dairy industry, whey, and oil mill waste effluent (Fig. 2).

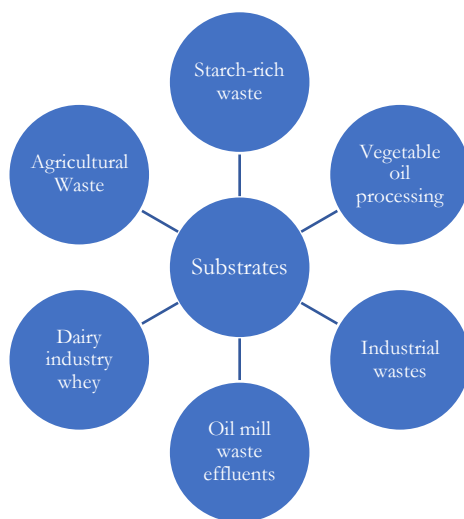


Figure 2 Substrates for biosurfactant production

Pre-treatment of substrates used for biosurfactant production

The substrates used for biosurfactant production can be pretreated by using acid such as hydrochloric acid (HCl), sulfuric acid (H₂SO₄), alkali such as sodium hydroxide (NaOH), potassium hydroxide (KOH), use of enzymes, drying, etc. (Fig. 3). The pre-treatment of the substrates will reduce the size of the substrates and make it suitable for biosurfactant production. Such pretreated substrates for the biosurfactant production will be useful in solid state fermentation for biosurfactant production.

The production of biosurfactant depends on carbon and nitrogen ratio. The commonly used media include minimal salt medium and usually the yield is in the range of 1.7 mg/ml (Phulpoto et al., 2020). However, the cost of laboratory media is a major constraint for large-scale production of biosurfactant. In order to reduce the cost and make the production economical, lowering the substrate/media cost is essential. At the same time, if the components of media are renewable, the production becomes eco-friendly. There are reports on reuse of agrowastes for biosurfactant production (Kourmentza, 2017; Rivera et al., 2019; Sharma et al., 2020). The present review consolidates renewable resources and proposes cost-effective processes for biosurfactant production.

In recent years, many researchers have shown the possibility of various agricultural, industrial wastes, etc. for utilization of biosurfactant production. They can be

categorized as fruit, vegetable, agricultural wastes, starch-rich, lignocellulose, industrial, dairy industry whey, vegetable oil, etc. In order to make the production eco-friendly and cost-effective, low cost substrates and reduced production time is desired along with minimum energy requirements for fermentation process and availability of resources.

Biosurfactant production using solid state fermentation

Biosurfactants can be produced by solid state fermentation (SSF) using economical and eco-friendly substrates or wastes such as fruit, vegetable, agricultural wastes, etc. SSF is fermentation in the absence or near absence of water. It is a biological process (carried out with the help of microorganisms). The process of biosurfactant production using SSF is represented in Fig. 4. The substrate is inoculated with the microorganism (count- 1x10⁸cells/ml) and then kept for SSF for 25 days. The leachate obtained is analyzed for biosurfactant production after every 5 days of time period.

Biosurfactant production using lignocellulosic wastes

Lignocellulose is a carbon source which is generated in huge quantities. They are composed of cellulose, hemicelluloses and lignin. There are reports on biosurfactant production using lignocellulosic wastes (Bezerra et al., 2019; Joy et al., 2019; Konishi et al., 2015; Panjar et al., 2020). The different lignocellulosic wastes



include bagasse, press mud, sugarcane trash, corn cob, orange, apple peel, coconut shell, rice, and wheat straw, etc.

et al., 2021; Kourmentza, 2017; Rivera et al., 2019; Sharma et al., 2020).

Fruit and vegetable waste for biosurfactant production

There are several reports where fruits and vegetable wastes such as apples, banana, orange peel, carrot, etc. are used as the best substrates for rhamnolipids biosurfactant (Chebbi

Food waste for biosurfactant production

Food waste can be used as substrate for biosurfactant production. The biosurfactant production using food waste as substrate is shown in Fig. 5.

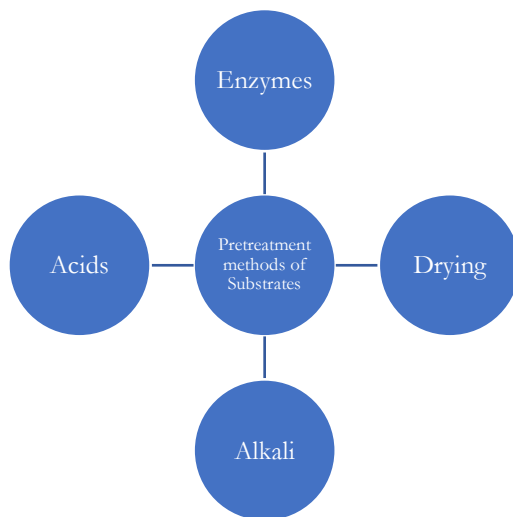


Figure 3 Pretreatment methods of substrates for biosurfactants production

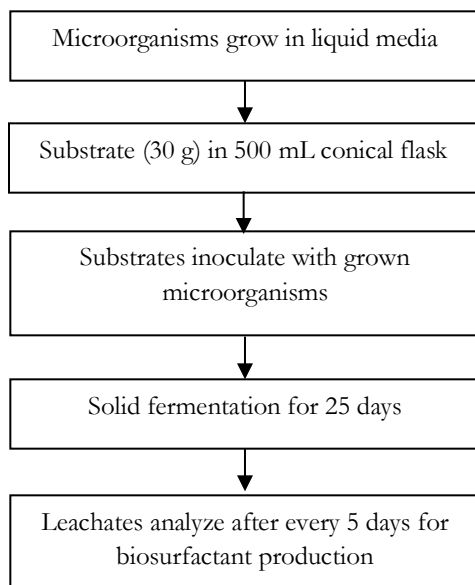


Figure 4 Biosurfactant production using solid state fermentation

Advantages of renewable sources for biosurfactant production

The advantages of various renewable sources for biosurfactant production are they are non-toxic, economical, and easily available in huge amounts. These renewable sources include molasses, cassava wastewater, animal fat, frying oil, cashew apple, glycerol, agro-industrial residues, whey, etc. (Fig. 6). The renewable substrates (%)

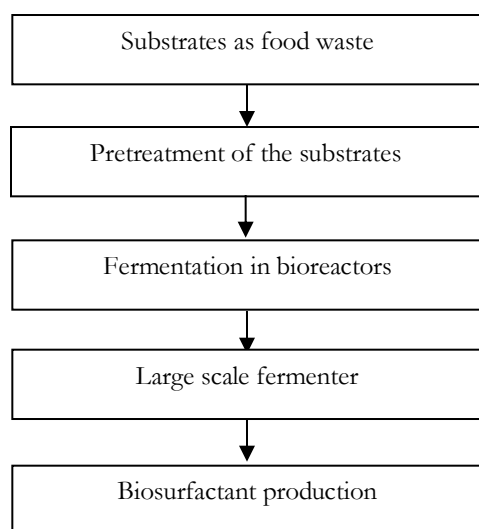


Figure 5 Biosurfactant production using food waste as substrates

for biosurfactant production (Raza et al., 2007) are shown in Fig. 6.

Substrates used for biosurfactant production

(Vandana & Singh, 2018)

Agricultural wastes

The agro-waste as substrate for biosurfactant production is shown in Fig. 7.

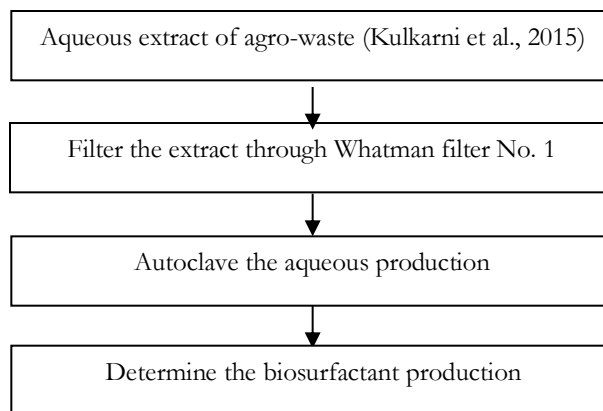


Figure 6 Renewable sources for biosurfactant production

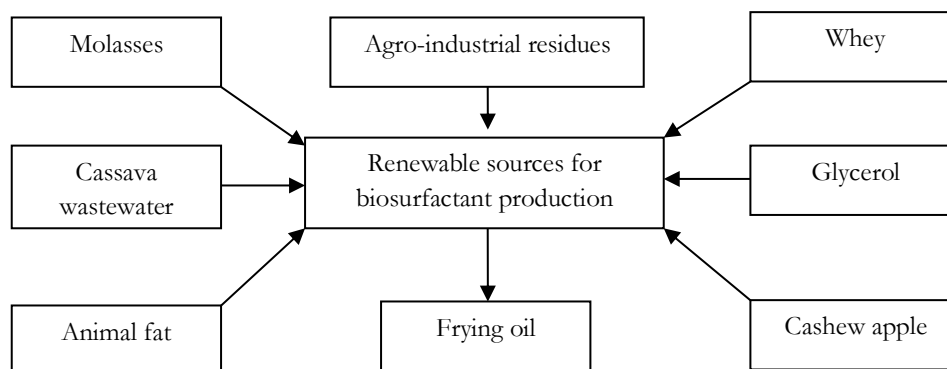


Figure 7 Agro-waste as substrate for biosurfactant production

Dairy industry whey

Curd whey is processed by removing casein and then deproteinized whey is used as substrate for biosurfactant production.

Industrial wastes

Bagasse, distillery waste, sugar industry effluent, fruit processing wastes is used as cheap substrates for biosurfactant production.

Vegetable oil

Vegetable oil is used as substrate for biosurfactant production.

Microorganisms producing biosurfactant

The microorganisms (bacteria, fungi, yeast, actinobacteria) producing biosurfactant are *Bacillus*, *Pseudomonas*, *Streptomyces*, *Acinetobacter*, *Candida*, *Aspergillus*, *Paenibacillus*,



Rhodococcus sp., *Serratia marcescens*, *Yarrowia lipolytica*, etc. The microorganisms and renewable substrates for biosurfactant production are represented in Table 1.

Microorganisms and biosurfactant yield

The bacteria and biosurfactant yield in represented in Table 2. And the fungi and biosurfactant yield in represented in Table 3.

Composition of various substrates used for biosurfactant production

a) Molasses: Dry matter - 76.8, Glucose - 5.29, phosphates - 2.03, potassium - 1.82, calcium - 1.39, magnesium - 0.43, total sugars - 62.3 (%), nitrates - 464 (mg/kg) (Palmonari et al., 2020).

b) Bagasse - Cellulose - 35.2, hemicelluloses - 24.5, lignin - 22.2, ash - 20.9 (%) (Rezende et al., 2011).

c) Pressmud - Nitrogen - 1.15-3.0, Phosphorus - 0.60-3.50, Potassium - 0.30-1.80, crude wax - 5-14, ash - 9-10, MgO - 0.5-1.5, CaO - 1-4 (%) (Rassppan et al., 2015).

d) Corn steep liquor - Carbohydrates - 5.8, proteins - 24, fats -1, minerals - 8.2 (%) (Joshi et al., 2018).

e) Vegetable oil - fatty acids

f) Rice straw - Cellulose - 49.29, hemicellulose - 11.55, silica - 17.07, organic matter - 80.08, ash - 19.92, crude protein - 4.40 (%)

Table 1 Microorganisms and renewable substrates for biosurfactant production

Microorganism	Renewable substrate	Reference
<i>Candida lipolytica</i>	Waste soybean oil and corn steep liquor	(Souza et al., 2016)
<i>Candida glabrata</i>	Corn steep liquor and whey	(Lima et al., 2017)
<i>Candida glabrata</i>	Corn steep liquor, whey and cassava wastewater	(Silva et al., 2015)
<i>Cunninghamella bertholletiae</i>	Waste soy oil and corn steep liquor	(Chebbi et al., 2021)
<i>Rhizopus arrhizus</i>	Soybean post-frying oil/sodium glutamate	(Pele et al., 2018)

Table 2 Bacteria and biosurfactant yield

Bacteria	Biosurfactant	Substrate	Yield (mg/l)	Reference
<i>Pseudomonas</i> sp.	Rhamnolipid		2070	(Priji et al., 2017)
<i>B. kauriensis</i> KP23	Rhamnolipid		780	(Tavares et al., 2013)
<i>Bacillus subtilis</i> ATCC 6051		Brewery waste (trub)	100.76	(Nazareth et al., 2021)
<i>Bacillus subtilis</i> MTCC 2423	Surfactin	Rice mill polishing residue	4.17	(Gurjar & Sengupta, 2015)
<i>Bacillus licheniformis</i> AL1.1	Lichenysin	Molasses	3.2	(Coronel-León et al., 2016)
<i>Bacillus pseudomycolides</i>	Lipopeptide	Soybean oil waste	56	(Li et al., 2016)
<i>Bacillus subtilis</i> DSM 3256	Surfactin	Olive mill waste	0.068	(Maass et al., 2016)
<i>Bacillus nealsonii</i> S2MT		Glycerol (2% v/v) and NH ₄ NO ₃ (0.1% w/v)	1300	(Phulpoto et al., 2020)
<i>Staphylococcus</i> sp.	Lipopeptides	Residual frying oil	65-750	(Hentati et al., 2021)
<i>Bacillus subtilis</i>		Corn steep liquor	1.3	(Gudina et al., 2015)
<i>Pseudomonas cepacia</i> CCT6659		Canola waste frying oil	40500	(Soares et al., 2018)
<i>Pseudomonas aeruginosa</i> M408		Olive oil	12600	(Ji et al., 2016)

Table 3 Fungi and biosurfactant yield

Fungi	Substrate	Yield	Reference
<i>Candida bombicola</i>	Corn steep liquor, molasses, soybean waste frying oil 5% (v/v)	61	(Pinto et al., 2018)
<i>Candida tropicalis</i> UCP0996	Sugarcane molasses, corn steep liquor, waste frying oil (2.5%)	7.36	(Almeida et al. 2017)
<i>Candida lipolytica</i> UCP 0988	Animal fat (5%) and corn steep liquor (2.5%)	40	(Santos et al., 2017)
<i>Candida bombicola</i> URM 3718	Molasses, corn steep liquor, soybean waste frying oil 5% (v/v)	8.4	(Luna et al., 2016)
<i>Candida sphaerica</i>	Ground-nut oil refinery residue (9%) and corn-steep liquor (9%)	21	(Luna et al., 2015)
<i>Aureobasidium pullulans</i>	Sucrose (50 gl ⁻¹), peptone (0.6 gl ⁻¹) and yeast extract (0.4 gl ⁻¹)	15	(Saur et al., 2019)
<i>Rhodotorula paludigena</i>	Glucose (150 gl ⁻¹) and yeast extract (1.5 gl ⁻¹)	20.90	(Garay et al., 2017)

Screening methods for biosurfactant production by microorganisms (Vandana & Singh D, 2018)

The different screening methods for biosurfactant production by microorganisms are represented in Fig. 8.

Drop collapse test

The drop collapse test is used to check if microorganisms are producing biosurfactant. In this method, a drop of oil is placed on a clean glass slide. To this, 10 μ l of microbial culture is added without disturbing the drop of oil. If the drop of oil is found to rupture in 1 min, it indicates production of biosurfactant by microorganism. There is a report on the study of rapid detection of biosurfactants by drop collapse assay on lotus leaf (*Nelumbo nucifera*) (Waghmode et al., 2016).

Blood agar hemolysis

Blood agar hemolysis is also of the methods to check for biosurfactant production by microorganism. On the blood agar medium, if the microorganism causes lysis of the blood cells, there is presence of a colorless ring around the colony which indicates the production of biosurfactant.

Oil displacement activity / oil spreading method

This method for screening of biosurfactant requires less volume of sample and is very fast. In this method, the biosurfactant if present, the oil will be displaced, and it will show presence of zone of clearance. The increase in diameter of the zone is calculated.

Emulsification index assay

Emulsification index assay is also one of the methods for the screening of biosurfactant production by microorganisms.

Cetyl trimethyl ammonium bromide (CTAB) agar plate method

This method is used to check for the production of anionic biosurfactants. In this method, the microorganisms if producing anionic biosurfactants, it forms blue colored insoluble ion which pairs with methylene blue. The presence of blue-colored halos around the colonies indicates production of anionic biosurfactants by microorganisms.

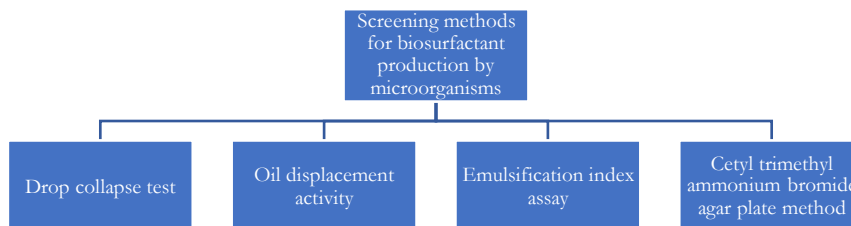


Figure 8 Screening methods for biosurfactant production by microorganisms

Factors affecting biosurfactant production

The media composition used for the growth of microorganisms plays an important role in biosurfactant production. Carbon (C) and nitrogen (N) sources are required for the growth of microorganisms. The C: N ratio affects biosurfactant production (Singh et al., 2019). In the media, the carbon sources used are viz., glucose, sucrose, and glycerol, while yeast extract, NaNO_3 , urea and soya broth as nitrogen sources for the growth of microorganisms (Wu et al., 2018). The carbon and nitrogen sources must be added in proper amount for maximum biosurfactant production. With glucose as carbon and sodium nitrate as nitrogen source, the rhamnolipid production was 5.28 and 4.38 g/l respectively (Onwosi & Odibo, 2012). There is a report on glycolipoprotein biosurfactant production by *Oceanobacillus* sp. BRI 10. The biosurfactant produced was higher (E24 = 55%) in the medium having glucose (3%) and ammonium chloride (0.48%) at 30 $^{\circ}\text{C}$, pH 8.0 after 48 h (Jadhav et al., 2013). There is a study where glucose in basal salt medium was replaced with sugarcane juice, whey and local commercial table sugar. The maximum emulsification index (E24) of 67.4% was obtained with sugarcane juice (Parhi et al.,

2016). The best nitrogen source found for biosurfactant production was sodium nitrate. E24 increased to 68.74% with sugarcane juice and sodium nitrate. The yield of biosurfactant achieved was 14.25 g l^{-1} with a 14-fold increase in the yield and eight times decrease in the production cost (Parhi et al., 2016).

Genetically engineered microorganisms for biosurfactant production

Genetically engineered microorganisms with optimized cultural conditions and other parameters will be effective to produce good quality biosurfactant. There is a need for research on expression of biosurfactant genes in hosts for improved and good quality production of biosurfactant. The molecular aspects combined with genetic engineering of biosynthetic genes are gaining attention of the researchers and this molecular aspect will improve the synthesis of biosurfactants which is having industrial, agricultural, etc. applications (Jimoh et al., 2021). The recombinants will improve the nature, quality and quantity of biosurfactants. The research on use of genetically engineered microorganisms for biosurfactant production will have wide applications in various industries.

Conclusions

Biosurfactants produced from renewable sources will be eco-friendly, economical, and easy. The biosurfactants produced will have many industrial applications. The substrates used for biosurfactant production are eco-friendly, economical, and easily available in huge amounts. The production of biosurfactants using microorganisms on a large-scale is very easy. The review is novel as it focuses on the use of renewable sources for biosurfactant production. More studies need to be carried out on genetically engineered microorganisms for biosurfactant production.

Acknowledgements

I would like to acknowledge Dr. Rama Bhadekar, Department of Microbial Biotechnology, Rajiv Gandhi Institute of IT and Biotechnology, Bharati Vidyapeeth (Deemed to be) University, Pune, Maharashtra, India for her suggestions in this review.

Conflict of Interest: The authors declare no conflict of interest.

Data Availability Statement: The data that support the finding of this study are available from the corresponding author, upon reasonable request.

References

- Akbari, S., Abdurahman, N., Yunus, R., Fayaz, F., & Alara, O. (2018). Biosurfactants - a new frontier for social and environmental safety: A mini review. *Biotechnology Research and Innovation*, 2, 81-90.
- Almeida, D., Soares da Silva, R., Luna, J., Rufino, R., Santos, V., & Sarubbo, L. (2017). Response surface methodology for optimizing the production of biosurfactant by *Candida tropicalis* on industrial waste substrates. *Frontiers in Microbiology*, 8, 157, 1-13.
- Bezerra, K., Gomes, U., Silva, R., Sarubbo, L., & Ribeiro, E. (2019). The potential application of biosurfactant produced by *Pseudomonas aeruginosa* TGC01 using crude glycerol on the enzymatic hydrolysis of lignocellulosic material. *Biodegradation*, 30, 351-361.
- Chebbi, A., Franzetti, A., Castro, F., Tovar, F., Tazzari, M., Scaffoni, S., & Vaccari, M. (2021). Potentials of winery and olive oil residues for the production of rhamnolipids and other biosurfactants: A step towards achieving a circular economy model. *Waste and Biomass Valorization*, 12, 4733-4743.
- Coronel-León, J., Marques, A., Bastida, J., & Manresa, A. (2016). Optimizing the production of the biosurfactant lichenysin and its application in biofilm control. *Journal of Applied Microbiology*, 120, 99-111.
- Fenibo, E., Douglas, S., & Stanley, H. (2019). Review on microbial surfactants: Production, classifications, properties and characterization. *Journal of Advances in Microbiology*, 18, 1-22.

- Garay, L., Sitepu, I., Cajka, T., Cathcart, E., Fiehn, O., German, J., Block, D., & Boundy-Mills, K. (2017). Simultaneous production of intracellular triacylglycerols and extracellular polyol esters of fatty acids by *Rhodotorula babjevae* and *Rhodotorula aff paludigena*. *Journal of Industrial Microbiology and Biotechnology*, 44, 1397-1413.
- Global Market Insights. (2018). Available at <https://www.gminsights.com/industry-analysis/biosurfactants-market-report>.
- Grand View Research. (2015). Available at <https://www.grandviewresearch.com/industry-analysis/biosurfactants-industry>.
- Gudina, E., Fernandes, E., Rodrigues, A., Teixeira, J., & Rodrigues, L. (2015). Biosurfactant production by *Bacillus subtilis* using corn steep liquor as culture medium. *Frontiers in Microbiology*, 6, 59, 1-7.
- Gurjar, J., & Sengupta, B. (2015). Production of surfactin from rice mill polishing residue by submerged fermentation using *Bacillus subtilis* MTCC 2423. *Bioresource Technology*, 189, 243-249.
- Hentati, D., Cheffi, M., Hadrich, F., Makhoulfi, N., Rabanal, F., Manresa, A., Sayadi, S., & Chamkha, M. (2021). Investigation of halotolerant marine *Staphylococcus* sp. CO100, as a promising hydrocarbon-degrading and biosurfactant-producing bacterium, under saline conditions. *Journal of Environmental Management*, 277, 111480. doi: 10.1016/j.jenvman.2020.111480.
- Jadhav, V., Yadav, A., Shouche, Y., Aphale, S., Moghe, A., Pillai, S., Arora, A., & Bhadekar, R. (2013). Studies on biosurfactant from *Oceanobacillus* sp. BRI 10 isolated from Antarctic sea water. *Desalination*, 318, 64-71.
- Ji, F., Li, L., Ma, S., Wang, J., & Bao, Y. (2016). Production of rhamnolipids with a high specificity by *Pseudomonas aeruginosa* M408 isolated from petroleum-contaminated soil using olive oil as sole carbon source. *Annals of Microbiology*, 66, 1145-1156.
- Jimoh, A., Senbadejo, Adeleke, R., & Lin, J. (2021). Development and genetic engineering of hyper producing microbial strains for improved synthesis of biosurfactants. *Molecular Biotechnology*, 63, 267-288.
- Joshi, S., Goyal, S., & Reddy, M. (2018). Corn steep liquor as a nutritional source for bio cementation and its impact on concrete structural properties. *Journal of Industrial Microbiology and Biotechnology*, 45, 657-667.
- Joy, S., Rahman, P., Khare, S., & Sharma, S. (2019). Production and characterization of glycolipid biosurfactant from *Achromobacter* sp. (PS1) isolate using one factor at a time (OFAT) approach with feasible utilization of ammonia soaked lignocellulosic pretreated residues. *Bioprocess and Biosystems Engineering*, 42, 1301-1315.
- Konishi, M., Yoshida, Y., & Horiuchi, J. (2015). Efficient production of sophorolipids by *Starmerella*



- bombicola* using a corncob hydrolysate medium. *Journal of Bioscience and Bioengineering*, 119, 317-322.
- Kourmentza, C., Freitas, F., Alves, V., & Reis, M. (2017). Microbial conversion of waste and surplus materials into high-value added products: The case of biosurfactants. In V. Kalia & P. Kumar (Eds.), *Microbial applications- Bioremediation and Bioenergy*. Springer International Publishing AG, pp. 29-77.
- Kulkarni, S., Kanekar, P., Jog, J., Sarnaik, S., & Nilegaonkar, S. (2015). Production of copolymer, poly (hydroxybutyrate-co-hydroxyvalerate) by *Halomonas campisalis* MCM B-1027 using agro-wastes. *International Journal of Biological Macromolecules*, 72, 784-789.
- Li, J., Deng, M., Wang, Y., & Chen, W. (2016). Production and characteristics of biosurfactant produced by *Bacillus pseudomycoloides* BS6 utilizing soybean oil waste. *International Biodeterioration & Biodegradation*, 112, 72-79.
- Lima, R., Rodriguez, D., Andrade, R., & Takai, G. (2017). Production and characterization of biosurfactant isolated from *Candida glabrata* using renewable substrates. *African Journal of Microbiology Research*, 11, 237-244.
- Luna, J., Rufino, R., Maria, A., Jara, A., Brasileiro, P., & Sarubbo, L. (2015). Environmental applications of the biosurfactant produced by *Candida sphaerica* cultivated in low-cost substrates. *Colloids and Surfaces A: Physico-chem and Engineering Aspects*, 480, 413-418.
- Luna, J., Santos, A., Rufino, R., & Sarubbo, L. (2016). Production of biosurfactant from *Candida bombicola* URM 3718 for environmental applications. *Chemical Engineering*, 49, 583-588.
- Maass, D., Moya, R., Garcia, R., Jurado, A., Ulson, A., Borges, J., & Altmajer D. (2016). Two-phase olive mill waste (alpeorujo) as carbon source for biosurfactant production. *Journal of Chemical Technology & Biotechnology*, 91, 1990-1997.
- Markets and Markets. (2016). Available at <https://www.marketsandmarkets.com/Market-Reports/biosurfactants-market-493.html>.
- Nazareth, T., Zanutto, C., Maass, D., de Souza, A. & Ulson, S. (2021). A low-cost brewery waste as a carbon source in biosurfactant production. *Bioprocess and Biosystems Engineering*, 44, 2269-2276.
- Onwosi, C., & Odibo, F. (2012). Effects of carbon and nitrogen sources on rhamnolipid biosurfactant production by *Pseudomonas nitroreducens* isolated from soil. *World Journal of Microbiology and Biotechnology*, 28, 937-942.
- Palmonari, A., Cavallini, D., Sniffen, C., Fernandes, L., Holder, P., Fagioli, L., Fusaro, I., Biagi, G., Formigoni, A., & Mammi, L. (2020). Characterization of molasses chemical composition. *Journal of Dairy Science*, 103, 6244-6249.
- Panjiar, N., Mattam, A., Jose, S., Gandham, S., & Velankar, H. (2020). Valorization of xyloserich hydrolysate from rice straw, an agro residue, through biosurfactant production by the soil bacterium *Serratia nematodiphila*. *Science of the Total Environment*, 729, 138933. doi: <https://doi.org/10.1016/j.scitotenv.2020.138933>.
- Parhi, P., Jadhav, V., & Bhadekar, R. (2016). Increase in production of biosurfactant from *Oceanobacillus* sp. BRI 10 using low cost substrates. *Songklanakarin Journal of Science and Technology*, 38, 207-211.
- Pele, M., Montero-Rodriguez, D., Ribeaux, D., Souza, A., Luna, M., Santiago, M., Andrade, R., Silva, T., Santiago, A., & Campos-Takak, G. (2018). Development and improved selected markers to biosurfactant and bioemulsifier production by *Rhizopus* strains isolated from Caatinga soil. *African Journal of Biotechnology*, 17, 150-157.
- Phulpoto, I., Yu, Z., Hu, B., Wang, Y., Ndayisenga, F., Li, J., Liang, H., & Qazi, M. (2020). Production and characterization of surfactin-like biosurfactant produced by novel strain *Bacillus nealsonii* S2MT and it's potential for oil contaminated soil remediation. *Microbial Cell Factories*, 19, 1-12.
- Pinto, M., Ribeiro, B., Guerrac, J., Rufino, R., Sarubbo, L., Santos, V., & Lunaa, J. (2018). Production in bioreactor, toxicity and stability of a low-cost biosurfactant. *Chemical Engineering*, 64, 595-600.
- Priji, P., Sajith, S., Unni, K., Anderson, R. & Benjamin, S. (2017). *Pseudomonas* sp. BUP6, a novel isolate from Malabari goat produces an efficient rhamnolipid type biosurfactant. *Journal of Basic Microbiology*, 57, 21-33.
- Rasspan, K., Kumar, A., & Santhosh, P. (2015). Studies on sugarcane press mud and distillery waste as a biofertilizer through composting. *International Journal of Chemical Science*, 13, 1333-1344.
- Raza, Z., Khan, M., & Khalid, Z. (2007). Physico-chemical and surface active properties of biosurfactant produced using molasses by a *Pseudomonas aeruginosa* mutant. *Journal of Environmental Science and Health: Toxic / Hazardous Substances and Environmental Engineering*, 42, 73-80.
- Rezende, C., de Lima, M., Maziero, P., deAzevedo, E., Garcia, W., & Polikarpov, I. (2011). Chemical and morphological characterization of sugarcane bagasse submitted to a delignification process for enhanced enzymatic digestibility. *Biotechnology for Biofuels*, 4, 54, 1-18.
- Rivera, A., Urbina, M., & YLopez, V. (2019). Advances on research in the use of agro-industrial waste in biosurfactant production. *World Journal of Microbiology and Biotechnology*, 35, 155, 1-18.
- Roy, A. (2017). Review on the biosurfactants: Properties, types and its applications. *Journal of Fundamentals of Renewable Energy and Applications*, 8, 248, 1-5.
- Santos, D., Rufino, R., Luna, J., Santos, V., & Sarubbo, L. (2016a). Biosurfactants: Multifunctional biomolecules of the 21st century. *International Journal of Molecular Sciences*, 17, 401, 1-31.



- Santos, D., Meira, H., Rufino, R., Luna, J., & Sarubbo, L. (2017). Biosurfactant production from *Candida lipolytica* in bioreactor and evaluation of its toxicity for application as a bioremediation agent. *Process Biochemistry*, 54, 20-27.
- Saur, K., Brumhard, O., Scholz, K., Hayen, H., & Tiso, T. (2019). A pH shift induces high-titer liamocin production in *Aureobasidium pullulans*. *Applied Microbiology and Biotechnology*, 103, 4741-4752.
- Sharma, P., Gaur, V., Kim, S., & Pandey, A. (2020). Microbial strategies for bio-transforming food waste into resources. *Bioresource Technology*, 299, 122580. doi: <https://doi.org/10.1016/j.biortech.2019.122580>.
- Singh, P., Patil, Y., & Rale, V. (2019). Biosurfactant production: Emerging trends and promising strategies. *Journal of Applied Microbiology*, 126, 2-13.
- Soares, S., de Almeida, D., Brasileiro, P., Rufino, R., de Luna, J. & Sarubbo, L. (2018). Production, formulation and cost estimation of a commercial biosurfactant. *Biodegradation*, 30, 191-201.
- Souza, P., Silva, T., Freitas-Silva, M., Andrade, R., Lima, M., Silva, P., Fonseca, T., & Campos-Takaki, G. (2016). Factorial design based medium optimization for the improved production of biosurfactant by *Mucor polymorphosphorus*. *International Journal of Current Microbiology and Applied Sciences*, 5, 898-905.
- Silva, G., Negreiros, J., Silva, N., Silva, T., Barbosa, R., Oliveira, N., & Okada, K. (2015). Campos-Takaki GM, Characterization of *Aspergillus niger* isolated from Caatinga soil with potential of biosurfactant production. In A. Mendez-Vilas (Ed), *Industrial, Medical and Environmental Applications of Microorganisms Current Status and Trends Madrid*, pp. 65.
- Tavares, L., Silva, P., Junqueira, M., Mariano, D., Nogueira, F., Domont, G., Freire, D., & Neves, B. (2013). Characterization of rhamnolipids produced by wild-type and engineered *Burkholderia kururiensis*. *Applied Microbiology and Biotechnology*, 97, 1909-1921.
- Vandana, P., & Singh, D. (2018). Review on biosurfactant production and its application. *International Journal of Current Microbiology and Applied Sciences*, 7, 4228-4241.
- Waghmode, M., Patil, N., Gaikwad, P., Gunjal, A., Nawani, N., & Kapadnis, B. (2016). Drop collapse assay on lotus leaf (*Nelumbo nucifera*): a simple and cost effective method for rapid detection of biosurfactants. *Journal of Experimental Biology and Agricultural Sciences*, 4, 505-511.
- Wu, T., Xu, J., Xie, W., Yao, Z., Yang, H., Sun, C., & Li, X. (2018). *Pseudomonas aeruginosa* L10: A hydrocarbon-degrading, biosurfactant-producing, and plant-growth-promoting endophytic bacterium isolated from a reed (*Phragmites australis*). *Frontiers in Microbiology*, 9, 1087, 1-12.