

# ANTIBIOTIC SENSITIVITY PATTERN OF BACTERIAL ISOLATES FROM CLINICAL SPECIMENS OF PATIENTS ADMITTED IN INTENSIVE CARE UNIT

Khilasa Pokharel<sup>1\*</sup>, Bishwa Raj Dawadi<sup>2</sup>

## Affiliation

1. Assistant Professor, Department of Microbiology, Kathmandu Medical College and Teaching Hospital, Nepal
2. Consultant, Emergency Medicine, Grande International College, Nepal

## ARTICLE INFO

Received : 27 July, 2021

Accepted : 22 December, 2021

Published : 21 February, 2022

© Authors retain copyright and grant the journal right of first publication with the work simultaneously licensed under Creative Commons Attribution License CC - BY 4.0 that allows others to share the work with an acknowledgment of the work's authorship and initial publication in this journal.



ORA 276

DOI: <https://doi.org/10.3126/bjhs.v6i3.43201>

## \* Corresponding Author

Dr. Khilasa Pokharel

Assistant Professor

Department of Microbiology

Kathmandu Medical College and Teaching Hospital, Nepal

Email: [khilaspokharel1@gmail.com](mailto:khilaspokharel1@gmail.com)

ORCID: <https://orcid.org/0000-0002-3683-3050>

## Citation

Antibiotic Sensitivity Pattern of Bacterial Isolates from Clinical Specimens of Patients Admitted in Intensive Care Unit. Khilasa Pokharel, Bishwa Raj Dawadi. BJHS 2021;6(3)16. 1601 - 1605.

## ABSTRACT

### Introduction

Antibiotics resistance is increasing nowadays. Factor to antibiotic resistance is its misuse. There is no national guideline for use of antibiotics.

### Objective

Study was conducted to determine bacteriological profile, and the bacterial antibiotic sensitivity pattern of the pathogens isolated from patients in intensive care unit.

### Methodology

Study was conducted between 1<sup>st</sup> October 2020 and 30<sup>th</sup> March 2021. The samples collected from Intensive care units patients were processed in Microbiology Laboratory following Clinical and Laboratory Standard Institute guideline. Organisms were identified morphologically and biochemically and antibiotic sensitivity pattern was determined by disc diffusion methods.

### Result

Out of total 397 samples, only 46 were found to be culture positive, of which 32 (69.56%) were gram negative organisms and 14 (30.43%) were gram positive organisms. Out of gram negative organisms *Klebsiella* species 10 (21.73%) were predominant which is more sensitive to colistin, polymyxin B and tigecycline. Among gram positive organisms *Staphylococcus aureus* 12 (85.71%) was found more sensitive to Vancomycin than others.

### Conclusion

It is concluded that *Klebsiella* species and *Staphylococcus aureus* were predominant pathogens isolated from the patients. Regular surveillance of antibiotic sensitivity pattern and stewardship program will be useful in treating patients in Intensive care unit.

## KEYWORDS

Antibiotics susceptibility, bacterial pathogens, intensive care unit.



## INTRODUCTION

Microbes are the organism that causes infections.<sup>1</sup> The major cause of mortality worldwide is infection which targeted low and lower middle countries.<sup>2</sup> Infection is common among hospitalized patients of intensive care unit.<sup>3,4</sup>

Intensive care unit (ICU) is a confined setting where antibiotics are extremely used.<sup>5</sup> Rather than other hospital patients, Intensive care unit patients are 5-10 times more likely of acquiring hospital infection.<sup>6</sup> Health care associated infection is high risk factor with critically ill patients in Intensive Care Unit which occurs due to various factors like decreased host defences, use of devices and due to cross transmission of infection among patients and staffs.<sup>7,8</sup> Health care associated infection and antibiotic resistance are prime treats among patients in ICU.<sup>9</sup> Therapeutic drugs are constantly found to be ineffective in infection, which is decreasing the success of routine treatment.<sup>10</sup> One study conducted in 2007 with ICU patients, from 75 different countries, reported that patients with longer ICU stay got higher rate of infection especially due to resistant *Staphylococci*, *Acinetobacter*, *Pseudomonas* species and *Candida* species.<sup>11</sup> ICU patients acquiring infections are mostly associated with invasive devices and ventilators.<sup>12</sup> The major consequences of the problem is increase in patients morbidity, mortality which is the result of the treatment failure.<sup>13,14</sup> Infections caused by gram negative bacteria in ICU has increased with which lack of treatment options against multi drug resistant (MDR) strains is in doubt. Infection that is caused by MDR gram negative organisms brings high morbidity and mortality. So, infection control guideline should be followed to improve patient outcome and decrease cost for treatment.<sup>15</sup> Antimicrobial Stewardship programs can optimize the proper use of available antimicrobial agent which can improve infections caused by organisms.

The results from this study can help in building up strategies for proper use of prophylactic and empiric antibiotic therapy among ICU patients in developing countries like ours. The aim of the study was to reduce the advance consequence of antimicrobial use.

## METHODOLOGY

This is a descriptive cross sectional study conducted at Kathmandu Medical College and Teaching Hospital from 1<sup>st</sup> October 2020 to 30<sup>th</sup> March 2021. Ethical approval was taken from Institutional review board (IRC) Reference no: 170920205. Clinical specimens collected from ICU patients were included in this study. Specimens after been collected in the Microbiology laboratory were further been processed for Grams staining and culture. After isolating the organisms from the culture they were further characterized by conventional biochemical test to identify the specific microorganism. Antibiotic susceptibility test of the bacterial isolates were done by Kirby Bauer disc diffusion method on Mueller Hinton Agar (MHA) as per Clinical Laboratory Standard Institute (CLSI) guideline. With a sterile loop four or

five colonies of the isolated cultured organisms were mixed with 2ml of sterile saline and was vortexed to create a suspension. Turbidity of the solution was adjusted to 0.5 Mc Farland standard. A sterile swab stick was then dipped into the formed suspension, firmly pressed to remove excess fluid and then lawn culture was performed on Muller Hinton agar (MHA). Antibiotic discs were then applied to MHA plates and was included at 37°C for 18-24 hours. Zone of inhibition was measured and interpreted using standard chart and organisms were reported susceptible, intermediate or resistant accordingly. Himedia, Mumbai, India and MAST diagnostics, Merseyside, England antibiotic discs were obtained.<sup>16</sup> Antibiotics of following concentrations used were: Amikacin (30µg), Gentamycin (10µg), Ciprofloxacin (5µg), Chloramphenicol (30µg), Co-Trimoxazole (25µg), Ceftazidime (30µg), Piperacillin-Tazobactam (100/10µg), Imipenem (10µg), Meropenem (10µg), Azithromycin (15µg), Penicillin (10µg), Oxacillin (1µg), Vancomycin (5µg), Linezolid (30µg), Tigecycline (30µg), Polymyxin B (30 units) and Colistin Sulphate (10µg) from Hi-Media laboratory. Zone diameter were interpreted according to Clinical and Laboratory standard Institute (CLSI 2017) recommendations.

Quality control for culture plates and antibiotic susceptibility were performed using *Escherichia coli* ATCC 25922, *Staphylococcus aureus* 25923, *Pseudomonas aeruginosa* ATCC 27853.

Multi Drug Resistance (MDR) bacterial isolates were identified according to the criteria recommended by international expert committee of the European Centre for disease Prevention and Control (ECDC) and the Centers for disease control and Prevention (CDC). The isolates resistant to at least one antimicrobial from three different group of first line drugs tests were regarded as Multidrug Resistant (MDR).<sup>17</sup>

Sample size (n) =  $Z^2 \alpha / 2 * (p) * (1-p) / d^2 = 366.23 \sim 367$

Where: Z = degree of confidence level = 95% = 1.96

p = prevalence = 60.8% = 0.608<sup>18</sup> d = allowable error = 5%

## RESULTS

During the study period a total of 397 samples were processed of which 46 (11.58%) were culture positive. Of total positive culture specimens higher number was isolated from pus 20 (43.47%), followed by fluid 9 (19.56%), urine 8 (17.39%), blood 5 (10.86%), catheter tip 2 (4.43%), endotracheal tip 1 (2.17%) and high vaginal swab 1 (2.17%). (Table No. 1)

**Table 1: Clinical manifestations**

Sample	Bacterial Isolate (%)
Pus	20 (43.47%)
Fluid	9 (19.56%)
Urine	8 (17.39%)
Blood	5 (10.86%)
Catheter tip	2 (4.34%)
Endotracheal tip	1 (2.17%)
High Vaginal Swab	1 (2.17%)
<b>Total</b>	<b>46 (100%)</b>



Among the bacterial isolates isolated from intensive care unit *Klebsiella* species was predominant among gram negative bacteria 10 (31.25%) and was mostly isolated from urine 5 (50.00%). Among gram positive organisms predominant was *Staphylococcus aureus* 12 (85.71%) and was mostly isolated from pus (91.66%).(Table No. 2)

microbiologist is required to get a better outcome.<sup>19</sup> Intensive care unit is common for secondary infection as compared to primary infection. In this study we have taken total 397 samples of which only 46 (11.58%) were reported as culture positive. This study showed that most of the isolates were from Pus 20 (43.47%) samples, followed by

**Table 2:** Distribution of bacterial isolates in Clinical Samples

Organisms	Pus	Fluid	Urine	Blood	Catheter tip	Endotracheal tip	High vaginal swab
<i>S.aureus</i>	11 (91.66%)	-	-	-	1 (8.33%)	-	-
<i>Streptococcus Species</i>	-	-	-	-	-	-	1 (100%)
<i>Coagulase negative Staphylococcus</i>	-	-	-	1 (100%)	-	-	-
<i>Klebsiellasppecies</i>	-	2 (20%)	5 (50%)	2 (20%)	1 (10%)	-	-
<i>Pseudomonas aeruginosa</i>	4 (50%)	2 (25%)	1 (12.50%)	-	-	1 (12.50%)	-
<i>Acinetobacter species</i>	2 (28.57%)	4 (57.14%)	1 (14.28%)	-	-	-	-
<i>Enterobacter species</i>	-	1 (100%)	-	-	-	-	-
<i>Proteus species</i>	3 (100%)	-	-	-	-	-	-
<i>Citrobacterspecies</i>	-	-	1 (100%)	-	-	-	-
<i>Escherichia coli</i>	2 (100%)	-	-	-	-	-	-

Among gram negative organisms *Klebsiellasppecies* is predominant in intensive care unit which is more sensitive to antibiotics like Colistin, Polymyxin B and Tigecycline. Out of gram positive organisms the most predominant organism was *Staphylococcus aureus* which is more sensitive to Vancomycin. (Table No. 3)

Fluid 9 (19.56%), Urine 8 (17.39%), Blood 5 (10.86%), Catheter tip 2 (4.34%), Endotracheal tube (ET) 1 (2.17%) and High Vaginal Swab 1 (2.17%). This study is slightly different with the study conducted by other authors<sup>20</sup> which shows highest percentage of growth in Blood 48 (37.79%), followed by ET 40 (31.49), Urine 39 (30.70%), Sputum 17

**Table 3:** Antibiotic sensitivity pattern of patients given in percentage in Intensive care unit

Antibiotics	<i>Klebsiellasp</i> (%)	<i>Pseudomonas aeruginosa</i> (%)	<i>Acinetobactersps</i> (%)	<i>Citrobactersps</i> (%)	<i>Entero-bactersps</i> (%)	<i>Proteus sps</i> (%)	<i>E.coli</i> (%)	<i>S.aureus</i> (%)	CONS (%)	<i>Strep.sps</i> (%)
Amikacin	70.00	50.00	42.84	100.00	100.00	66.66	50.00	33.33	100.00	100.00
Cefexime	60.00	25.00	28.57	00.00	00.00	00.00	00.00	33.33	00.00	00.00
Ceftriaxone	40.00	50.00	28.57	00.00	00.00	33.33	00.00	58.33	00.00	00.00
Cotrimoxazole	30.00	12.50	42.85	00.00	00.00	66.66	00.00	41.66	00.00	00.00
Ciprofloxacin	40.00	25.00	57.14	00.00	00.00	66.66	50.00	41.66	00.00	00.00
Chloramphenicol	30.00	37.50	28.57	00.00	00.00	66.66	50.00	33.33	100.00	00.00
Imipenem	70.00	37.50	28.57	100.00	100.00	100.00	100.00	-	-	-
Meropenem	80.00	37.50	28.57	100.00	100.00	100.00	100.00	-	-	-
Piperacillin/Tazobactam	90.00	25.00	57.14	100.00	100.00	100.00	100.00	-	-	-
Azithromycin	30.00	00.00	00.00	00.00	00.00	33.33	50.00	33.33	100.00	100.00
Ceftazidime	50.00	37.50	42.85	00.00	00.00	33.33	100.00	-	-	-
Penicillin	-	-	-	-	-	-	-	33.33	100.00	100.00
Colistin	100.00	100.00	100.00	100.00	100.00	100.00	100.00	-	-	-
Polymyxin B	100.00	100.00	100.00	100.00	100.00	100.00	100.00	-	-	-
Tigecycline	90.00	-	100.00	100.00	100.00	100.00	100.00	-	-	-
Gentamicin	-	-	-	-	-	-	-	41.66	100.00	100.00
Oxacillin	-	-	-	-	-	-	-	41.66	100.00	100.00
Vancomycin	-	-	-	-	-	-	-	100.00	-	100.00
Linezolid	-	-	-	-	-	-	-	91.66	100.00	-

## DISCUSSION

Antimicrobial resistance in the hospital infection is increasing day by day mostly when the infection is caused by multidrug resistance organisms. Intensive care unit is led by a team which involves intensivist, clinical microbiologist associated with medical and surgical specialities. In Intensive care unit coordination between the intensivist and clinical

(13.38%), Pus 11 (8.66%), Catheter 4 (3.14%), Ear Swab 2 (1.57%) and Stool 1 (0.78%). In this study, we isolated gram negative bacteria 32 (69.56%) as compared to gram positive cocci 14 (30.43%). In this study, we isolated more gram negative bacteria 32 (69.56%) compared to gram positive cocci 14 (30.43%). In the Canadian National ICU cases the result observed was *Pseudomonas aeruginosa*,



*Staphylococcus aureus*, *Haemophilus influenzae*, *Enterococcus* species and *Klebsiella pneumoniae*.<sup>21</sup> Most of the Asian countries, the pathogens that are mostly isolated from ICU patients were *Pseudomonas aeruginosa*, *Klebsiella* species, *Escherichia coli*, *Enterococcus* species and *Staphylococcus aureus*<sup>22</sup> which is similar to our study. Among all admitted cases who were culture positive gram negative organisms most of the infection was caused by *Klebsiella* species 10 (31.25%), followed by *Pseudomonas aeruginosa* 8 (25%), *Acinetobacter* species 7 (21.87%), *Proteus* 3 (9.37%), *Escherichia coli* 2 (6.25%), *Enterobacter* species 1 (3.12%) and *Citrobacter* species 1 (3.12%). This study is nearly similar to the study which show predominance of *Acinetobacter* species 52 (41%) followed by *Klebsiella pneumoniae* 36 (28%) and *Pseudomonas aeruginosa* 27 (21%) among ICU cases.<sup>23</sup> This study revealed result as the study investigated in ICU of Indonesia which shows predominance of *Pseudomonas aeruginosa*, *Klebsiella* species and *Escherichia coli*.<sup>24</sup> We found Enterobacteriaceae more sensitive to piperacillin/tazobactam and *Pseudomonas aeruginosa* more resistant to ceftazidime and more sensitive to imipenem. Similar type of study was conducted with the result showing Enterobacteriaceae showing 26.1% resistance to piperacillin/tazobactam and 64.1% of *Pseudomonas aeruginosa* resistant to ceftazidime and 42.0% to imipenem.<sup>25</sup> Our study shows that gram negative bacteria were resistant to commonly used antibiotics and were more sensitive to colistin, polymyxin B and tigecycline which is similar with other studies.<sup>26</sup> In this study Gram positive organism like *Staphylococcus aureus* is more sensitive to Vancomycin that is similar to other studies done<sup>27</sup> which shows predominance of *Staphylococcus aureus* in hospital setting along with its sensitivity toward Vancomycin.

Our result have got clinical importance in the treatment and management among intensive care unit patients. The study was conducted among ICU patients. For better clinical step empirical antibiotics with antibiotic stewardship program, which can prevent multidrug resistance and drug resistant

organism. We have also focused on using sterile techniques while inserting devices, use of sterile gloves and gowns in ICU and most importantly hand hygiene to prevent nosocomial infections and better patient response. We have analyzed the spectrum of pathogens and the antibiotic treatment.

## CONCLUSION

*Klebsiella* species, *Pseudomonas aeruginosa* and *Acinetobacter* species are the gram negative multiresistant organisms isolated. Most of the strains of these organism were resistant to ceftriaxone, cotrimoxazole, ciprofloxacin, chloramphenicol, ceftazidime and were more sensitive to colistin and polymyxin B. Among gram positive organisms *Staphylococcus aureus* was the pathogen prevalent in ICU and the strains were found to be more sensitive to vancomycin. Appropriate antibiotic utilization in ICU is required for which stewardship program is useful in ICU.

## LIMITATIONS OF STUDY

Antibiotic sensitivity test, its use in the hospital and hospital infection control vary from hospital to hospital. The limitation of this study is that we would have conducted study in large size but we conducted this study in only one hospital site and could not compare it with other hospital setting.

## ACKNOWLEDGEMENT

Thankful to all the staff of Microbiology Department working in our hospital for their support and active participation in this article.

## CONFLICT OF INTEREST

There was no conflict of interest during the study.

## FINANCIAL DISCLOSURE

There is no financial support in this study.

## REFERENCES

- Chaudhry D, Prajapat B. Intensive care unit bugs in India: How do they differ from the western world? The Journal of association of chest physicians. 2017;5(1):10-7. DOI: 10.4103/2320-8775.196645
- Lozano R, Naghavi M, Foreman K, Lim S, Shibuya K, Aboyans V et al. Global and regional mortality from 235 causes of death for 20 age groups in 1990 and 2010: a systematic analysis for the Global Burden of Disease Study 2010. Lancet. 2012 ;380(9859):2095-128. DOI: 10.1016/S0140-6736(12)61728-0
- Hanberger H, Garcia-Rodriguez JA, Gobernado M, Goossens H, Nilsson LE, Struelens MJ. Antibiotic susceptibility among aerobic gram-negative bacilli in intensive care units in 5 European countries. French and Portuguese ICU Study Groups. JAMA. 1999;281(1):67-71. DOI: 10.1001/jama.281.1.67
- Vincent JL, Bihari DJ, Suter PM, Bruining HA, White J, Nicolas-Chanoine MH, Wolff M, Spencer RC, Hemmer M. The prevalence of nosocomial infection in intensive care units in Europe. Results of the European Prevalence of Infection in Intensive Care (EPIC) Study. EPIC International Advisory Committee. JAMA. 1995;274(8):639-44. DOI: 10.1001/jama.281.1.67
- Marin H, Kollef MD, Victoria J, Fraser MD. Antibiotic Resistance in the Intensive Care Unit. Annals of Internal Medicine Reviews. 2001; 134(4):298-314. DOI: 10.7326/0003-4819-134-4-200102200-00014
- Weber DJ, Raasch R, Rutala WA. Nosocomial infections in the ICU: the growing importance of antibiotic-resistant pathogens. Chest. 1999;115(3):34-41. DOI: 10.1378/chest.115.suppl\_1.34s
- Trilla A. Epidemiology of nosocomial infections in adult intensive care units. Intensive Care Med. 1994;20(3):1-4. DOI: 10.1007/BF01745243
- Mihaly V, Orsolya B, Monica O, Anna PA, Hajna K, Maria CS. The Incidence and Risk Factors of Nosocomial Infections in ICU. Acta Medica Marisensis. 2016;62(3):304-8. DOI: 10.4103/0972-5229.148633
- Parajuli NP, Acharya SP, Mishra SK, Parajuli K, Rijal BP, Pokhrel BM. High burden of antimicrobial resistance among gram negative bacteria causing healthcare associated infections in a critical care unit of Nepa. J Pathog. 2017;6:67. DOI: http://dx.doi.org/10.3126/ mjsbh.v19i2.27689





10. Laxminarayan R , Duse A , Wattal C, Zaidi AKM, Wertheim HFL, Sumpradit N. Antibiotic resistance—the need for global solutions. The Lancet Infectious Diseases Commission. 2013 November;13(12):1057-98. DOI: 10.1016/S1473-3099(13)70318-9
11. Vincent JL, Rello J, Marshall J, Silva E, Anzueto A, Martin CD, et al. et al. International study of the prevalence and outcomes of infection in intensive care units. JAMA. 2009;302(21):2323–9. DOI: 10.1001/jama.2009.1754
12. Shulman L, Ost D. Managing infection in the critical care unit: How can infection control make the ICU safe. Crit Care Clin. 2005;21:111–128. DOI: 10.1016/j.ccc.2004.10.002
13. Vasudevan A, Memon BI, Mukhopadhyay A, Li J, Tambyah PA. The costs of nosocomial resistant gram negative intensive care unit infections among patients with the systemic inflammatory response syndrome- a propensity matched case control study. Antimicrobial Resistance and Infection Control. 2015 February;4(1). DOI: <https://doi.org/10.1186/s13756-015-0045-8>
14. Morales E, Cots F, Sala M, Comas M, Belvis F, Riu M et al. Hospital costs of nosocomial multi-drug resistant *Pseudomonas aeruginosa* acquisition. BMC Health Services Research. 2012;12(1):122. DOI: <https://doi.org/10.1186/1472-6963-12-122>
15. Paterson DL, Ko WC, Von Gottberg A, Mohapatra S, Casellas JM, Goossens H, et al. Antibiotic therapy for Klebsiellapneumoniae bacteremia: implications of production of extended-spectrum beta-lactamases. Clin Infect Dis. 2004;39(1):31–7. DOI: 10.1086/420816.
16. Clinical and Laboratory Standards Institute, “Performance standards for antimicrobial susceptibility testing; nineteenth informational supplement,” CLSI Document M100-S19, Clinical and Laboratory Standards Institute, Wayne, Pa, USA, 2009. ISBN Number: 978-1-68440-105-5
17. Magiorakos AP, Srinivasan A, Carey RB. Multidrug-resistant, extensively drug-resistant and pandrug-resistant bacteria: an international expert proposal for interim standard definitions for acquired resistance. Clinical Microbiology and Infection. 2012;18(3):268–81. DOI: 10.1111/j.1469-0691.2011.03570.x
18. Bhandari P, Thapa G, Pokhrel BM, Bhatta DR, Devkota U. Nosocomial Isolates and Their Drug Resistant Pattern in ICU Patients at National Institute of Neurological and Allied Sciences. International Journal of Microbiology. 2015. DOI: <https://doi.org/10.1155/2015/572163>
19. RK Sanjana, PC Majhi. Microbial infection and antibiotic patterns among intensive care unit patients in a tertiary hospital in Central Nepal. Journal of College of Medical Sciences-Nepal. 2012;8(3);1-8. DOI: 10.3126/jcmsn.v8i3.8678
20. Savanur SS, Gururaj H. Study of Antibiotic Sensitivity and Resistance Pattern of Bacterial Isolates in Intensive Care Unit Setup of a Tertiary Care Hospital. Indian Society of Critical Care Medicine. 2019;23(12):547-55. DOI: 10.5005/jp-journals-10071-23295
21. Zhanel GG, DeCorby M, Laing N, Weshnoweski B, Vashisht R, Taylor F, et al. Antimicrobial-resistant pathogens in intensive care units in Canada: results of the Canadian National Intensive Care Unit (CAN-ICU) study, 2005-2006. Antimicrob Agents Chemother. 2008;52(4):1430–7. DOI: 10.1128/AAC.01538-07
22. Radji M, Fauziah S, Aribinuko N. Antibiotic sensitivity pattern of bacterial pathogens in the intensive care unit of Fatmawati hospital. Indonesia Asian Pac J Trop Biomed. 2011;1(1):39–42. DOI: 10.1016/S2221-1691(11)60065-8
23. Siwakoti S, Subedi A, Sharma A, Baral R, Bhattarai NR, Khanal B. Incidence and outcomes of multidrug-resistant gram-negative bacteria infections in intensive care unit from Nepal- a prospective cohort study. Antimicrob Resist Infect Control. 2018;7:114. DOI: 10.1186/s13756-018-0404-3
24. Refdanita, Endang P, Nurgani A, Radji M. The sensitivity pattern of microorganisms against antibiotics at the Intensive Care Unit of Fatmawati Hospital Jakarta 2001-2002. J Makara. 2004;8(2):41–8. DOI: 10.1016/S2221-1691(11)60065-8
25. Mehta A, Rosenthal VD, Mehta Y, Chakravarthy M, Todi SK, Sen N et al. Device-associated nosocomial infection rates in intensive care units of seven Indian cities: findings of the International Nosocomial Infection Control Consortium (INICC) J Hosp Infect. 2007;67:168–74. DOI: 10.1016/j.jhin.2007.07.008
26. Bhandari P, Thapa G, Pokharel BM, Bhatta DR, Devkota U. Nosocomial isolates and their drug resistance pattern in ICU patients at National institute of Neurological and allied sciences, Nepal. Int J Microbiology. 2015;1-6. DOI: <https://doi.org/10.1155/2015/572163>
27. Bhattarai P, Dhungel BA, Shah P, Amatya J. Prevalence of Staphylococcus aureus in Intensive care Units and Post Operative Ward as a possible source of Nosocomial infection: An experience of tertiary care hospital. Janaki Medical College Journal of Medical Science. 2013;1(1):21-5. DOI: <https://doi.org/10.1155/2015/572163>

