

Etiology and Control of Citrus Canker Disease in Kavre

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Abstract

Citrus canker, caused by the bacterium *Xanthomonas citri* (Hase) occurs in large areas of the world's citrus growing countries including Nepal. Though the disease has serious effect in Nepal, this is the first detailed study carried out to isolate the pathogen and confirm it by available biochemical tests and pathogenicity test. Furthermore, the study was intended to find the proper and economical control measure to combat disease in citrus orchards. The causative agent of the disease was isolated from the diseased plants and pure culture was obtained. The isolated pure culture was subjected to Gram staining, catalase test, oxidase test, O-F test, starch hydrolysis, nitrate reduction test, methyl-red test, Voges-prausker test, indole production test, urease test and carbohydrate utilization test. To re-confirm it, pathogenicity test was conducted on host plant and after the appearance of the typical citrus canker lesion on host, the bacteria was re-isolated, thus proving the Koch's postulates. Different controlling chemicals, copperoxychloride (2.5%), copperoxychloride + kasugamycin (1000X), bordeaux mixture 1% and 2% were sprayed to the plants in citrus orchard at Dhulikhel and the decrease in disease severity after spraying of the chemicals was calculated with reference to the plants that were not sprayed with the chemicals. It was observed that spraying of the chemicals help in decreasing the disease severity. The chemical spray however was not able to eradicate the disease. The study concluded that *Xanthomonas citri* was the causative agent of the disease citrus canker and copper based chemicals when sprayed very early with the appearance of first symptoms of the disease could eliminate it in citrus fruits to minimum level.

Key words: Copper fungicides, citrus canker, Lime, *Xanthomonas citri*, acid lime, *Citrus aurantifolia*

Introduction

In the fiscal year 2004/2005, the total area of citrus cultivation was 25,909 ha out of which 14,606 ha was described as productive area with a total production of 15,956 mt (10.75mt./ha) of fruits. Lime is one of the citrus fruits cultivated in Nepal. In the year 2004/2005, the total production of lime was 19,132 mt (HMG/N, 2004/2005).

Of all the agricultural pests and diseases that threaten lime, citrus canker is one of the most devastating. The disease caused by bacterium *Xanthomonas citri* (Hase) occurs in large areas of the world. The disease is endemic in India, Japan and other south-East Asian countries, from where it has spread to all other citrus producing continents except Europe. Considering the high demand of lime in local markets that is being fulfilled

by import from India Government of Nepal has initiated Citrus (Lime) Mission Programme in Tehrathum, Bhojpur and Dhankuta districts with an objective to substitute the import of this fruits by increasing its production. No detail research has been carried out in Nepal on citrus canker. This study was carried out during 2005 to 2006 in Kavrepalanchowk district as it is representative of most of the hilly districts of Nepal.

Regarding the pathogen of citrus canker, in the late 1980s, strains associated with canker A were proposed as a new species, *Xanthomonas citri*, whereas types B and C as well as other strains causing citrus bacterial spot remained within *X. campestris* as pathovars *aurantifolii* and *citrumelo* respectively (Gabriel *et al.* 1989). Schaad *et al.* (2000) proposed a

reclassification that places citrus canker and citrus bacterial spot strains within *Xanthomonas* as a species *citri* (A strains), *aurantifolli* (B and C strains) and *citrumelo* (citrus bacterial spot strains). However, others authors rejected this new proposal, citing insufficient data to justify the removal of these strains from the species *axonopodis* (Vauterin *et al.* 2000, Young *et al.* 2001). Most recently, Brunings and Gabriel (2003) proposed the retention of *X. citri* as a species that includes only citrus canker strains (A and B-C)

Materials and Methods

Survey

Survey on canker infestation was carried out in different citrus orchards of Sarada Batase, Khanal Thok and Dhulikhel of Kavrepalanchowk district. In the survey, Citrus plants were checked for the symptoms which include raised, corky, tan lesions with water-soaked margins and yellow halos on the leaves

Disease Status and its severity

Status of the disease was found out by calculating disease frequency and severity. Disease frequency and severity were calculated by using the following formulae. (Johnston & Booth 1983)

$$\text{Disease frequency} = \frac{\text{Number of infected plants}}{\text{Total no. of plants}} \times 10$$

Severity of disease of each sampled plant was calculated by counting the number of leaves showing canker lesion out of total number of leaves sampled.

$$\text{Disease severity (\%)} = \frac{\text{No. of leaves with lesion}}{\text{Total no. of leaves}} \times 100$$

Study of pathogen

Laboratory works were conducted at Nepal Academy for Science and Technology (NAST), Khumaltar, Lalitpur. Leaf samples used for isolation of the bacteria were collected from an orchard of Dhulikhel. Using a sterile razor blade, younger portions of the lesion was cut from a recently collected material. Then a leaf with lesion was taken on a clean microscopic slide and a drop of sterile water was added and observed under oil

immersion objective to see streaming bacteria from the edge of leaf. With this characteristic, the isolation procedure was initiated. The cut portion was surface sterilized using 0.1% mercuric chloride and then washed repeatedly with sterile distilled water, transferred to another sterile plate, minced with a flamed razor in 1 ml of sterile distilled water. It was then allowed to stand for 10-15 min, and then serially diluted up to 10⁻³ dilution. Each diluent (0.2) was then spread on the plates on glucose yeast chalk agar (GYCA). The plates were then incubated at 28 °C for 72 hrs. Then suspected mucoid colonies with typical yellow pigment were subjected to Gram staining followed by biochemical and physiological tests. The biochemical tests performed were catalase test, oxidase test, oxidative-fermentative test, methyl Red and Voges-Proskauer tests, indole test, nitrate reduction test and urease test. Similarly carbohydrate utilization test was performed (carried on Dye's medium C) using carbohydrates; arabinose, mannose, galactose, glucose, sucrose, fructose, and sorbitol with incubation of 21 days at 27 °C. For the pathogenicity test, disease free lime saplings were pot cultured. Five plants were assigned for different tests and one plant was assigned for control. Single isolated colony from the 3 day old culture in GYCA was taken and mixed with sterile distilled water to prepare bacterial suspension containing 10⁶-10⁷ CFU/ml (compared with Mac Ferland's Scale at the suspension at 0.1 to 0.2 ml was inoculated on 3 to 4 points of each leaf. In each test plant 2 to 3 leaves were inoculated with bacterial suspension using 1ml syringe. Similarly sterile water was inoculated to the plant which served as a control. Inoculated plants were then incubated. Symptoms were observed on leaves after four days of inoculation.

Study of control measures

For the control of the disease, 5 plots i.e. 1st, 2nd, 3rd, 4th and 5th, with five plants in each plots (P1, P2, P3, P4 and P5) were treated with copper oxychloride (2.5%), copper oxychloride + Kasugamycin (1000X), Bordeaux mixture (1%), Bordeaux mixture (2%) and water (control) respectively. Altogether, 5 observations were done, out of which 2 were done before application of the treatments and 3 observations after the treatments of plants with

respective chemical sprays. Disease severity of the plants before and after each treatment was observed at 15 days interval.

Effectiveness of treatments

Effectiveness of treatments was determined on the basis of their capacity to reduce disease severity, which was calculated as follows (Table 3 and 4).

Effectiveness = Final disease severity (5th observation i.e. after 3rd treatment) - initial disease severity (1st observation).

Data Analysis

Effectiveness of treatments was compared by LSD (5%) test.

Table 1. Species of citrus plants infected with citrus canker in the surveyed area

Species of citrus plants	No. of plants with citrus canker /No. of plant observed		
	Dhulikhel	Khanalthok	Saradha Batase
<i>C. aurantifolia</i> (Acid lime)	48/55	9/9	0/0
<i>C. reticulata</i> (Mandarin orange)	0/0	0/15	0/72
<i>C. sinensis</i> (Sweet orange)	0/0	0/0	0/25

Results showed that only *C. aurantifolia* grown in Kavre were susceptible to citrus canker as other species such as *C. reticulata* and *C. sinensis* growing in the vicinity were uninfected. Thus our observation comply with the finding of Civerolo (1984), Graham (1992) and Stall (1993) who reported that acid lime is

Results and Discussion

Distribution

In the orchard of Dhulikhel, only *C. aurantifolia* was cultivated where out of total 55 plants, 48 were infected with different level of severity and 7 were found uninfected. In Khanalthok, out of 24 citrus fruits cultivated which included *C. aurantifolia* and *C. reticulata*, only 9 of *C. aurantifolia* were found infected, though both the fruits were grown in close vicinity. In Saradha Batase, a total of 97 citrus plants were observed which included *C. reticulata* and *C. sinensis* and none of them were infected with the disease citrus canker (Table 1.)

more susceptible to citrus canker than sweet orange and mandarin.

Confirmation of pathogens

The results of our study based on the morphological, microbiological and physiological characteristics (Table 2.) confirm that the pathogen was *X. citri*.

Table 2. Biochemical tests performed for confirmation of *X. citri*

S.No	Test performed	Results
1.	Gram staining	Gram negative rod
2.	Catalase test	Positive
3.	Oxidase test	Negative
4.	O-F test	Oxidative
5.	Methyl-Red test	Negative
6.	Voges- Proskeur test	Negative
7.	Indole test	Negative
8.	Urease test	Negative
9.	Strach hydrolysis test	Positive
10.	Nitrate reduction test	Negative
11.	Growth on nutrient agar incorporated with 0.1% triphenyl tetrazolium chloride	No growth observed
12.	Growth at 37°C	Positive
13.	Acid production from carbohydrates (arabinose, glucose, galactose, fructose, sorbitol)	Acid production from glucose, sucrose, mannose and galactose

Dynamics of disease development

Dynamics of the disease development was studied on the basis of increase in disease severity in control plots sprayed with water in every 15 days interval.

Efficiency of treatments

Statistical analysis of data (LSD, at 5% level of significance) (Table 3) showed that chemical sprays satisfactorily reduced the disease severity. Treatments had their effect immediately after use and continuously slowed down the disease development through out the rainy season as described by Koizumi (1985), Leite and Mohan (1990), Stall *et al.* (1980), (1982b) and Graham *et al.* (1992) who recommended copper-based bactericides as standard control measures for citrus canker world-wide. It was clearly observed that chemical treatments though effective could not control already existing lesions so the treatment should be started with the early sign of symptoms so that the disease spread can be slowed down or checked.

Copper hydroxide, basic copper chloride, copper oxychloride, and tribasic copper sulfate are the most effective bacterial sprays for protecting leaves and fruit from attack of *X. citri*. These materials can reduce the incidence of the disease, but they will not eliminate established infections. Copperoxychloride and kasugamycin have been used with satisfactory results. However, extensive use of copper may also cause phytotoxicity problems in treated groves (International citrus canker research workshop. 2000). The plants treated with Bordeaux mixture 2% showed symptoms of phytotoxicity. Therefore, Bordeaux mixture 2% may not be applicable to control the disease canker.

Table 3. Disease severity on first observation (2063/2/27)

Replicates	Plot 1	Plot 2	Plot 3	Plot 4	Plot 5
1	55%	50%	60%	70%	50%
2	60%	70%	60%	65%	70%
3	50%	65%	65%	60%	70%
4	58%	65%	70%	70%	65%
5	65%	75%	40%	60%	30%

(Observation after 3rd spray) (2063/5/15)

Table 4. Disease severity on 5th observation

Replicates	Plot 1	Plot 2	Plot 3	Plot 4	Plot 5
1	53%	51%	53%	72%	75%
2	57%	78%	61%	62%	85%
3	52%	67%	61%	61%	85%
4	64%	69%	73%	74%	82%
5	68%	75%	44%	57%	59%

Table 5. Mean effectiveness of each treatment

Effectiveness = Final disease severity (5th observation i.e. after 3rd treatment) - initial disease severity (1st observation)

Replicates	Plot1 Copperoxychloride (2.5%)	Plot 2 Copperoxychloride +kasugamycin (1000X)	Plot 3 Bordeaux (1%)	Plot 4 Bordeaux (2%)	Plot 5 Control (Water)	LSD*
1	-2	1	-7	2	25	
2	-3	8	1	-3	15	
3	2	2	-4	1	15	
4	6	4	3	4	17	
5	3	0	4	-3	29	
Mean effect iveness	1.2 ^b	3 ^b	-0.6 ^b	0.2 ^b	20.2 ^a	5.81

* at 5% level of significance

On the basis of statistical analysis (LSD, at 5%), it was found that all the treatments significantly reduced the rate of increase of the disease as compared to control.

The local varieties/cultivars of mandarin and sweet oranges were found resistant to citrus canker. The lime was very susceptible and special care has to be taken to control it with copper based chemical sprays.

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References

- Brunings, A.M. and D.W. Gabriel 2003. *Xanthomonas citri*: breaking the surface. *Molecular plant pathology* **4**: 141–157.
- Civerolo, E.L. 1984. Bacterial canker disease of citrus. *Journal of Rio Grande Valley Horticulture Society* **37**: 127-146.
- Graham, J.H., T.R. Gottwald, T.D. Riley and M.A. Bruce 1992. Susceptibility of citrus fruit to bacterial spot and citrus canker. *Phytopathology* **82**: 452-1325
- Gabriel, D.W., M.T. Kingsle, J.E. Hunter and T.R. Gottwald, T.R. 1989. Reinstatement of *Xanthomonas citri* (ex Hasse) and *X. phaseoli* (ex Smith) to species and reclassification of all *X. campestris* pv. *citri* strains. *International Journal of Systemic Bacteriology* **39**: 14–22.
- Johnston, A. and C. Booth. 1983. *Plant Pathologists Pocketbook (2nd ed)*. Commonwealth Mycological Institute, Kew, Surrey, England pp. 23-28,
- Koizumi, M. 1985. Citrus canker: The world situation. In: *Citrus Canker: An International Perspective* (Ed. L.W. Timmer), University of Florida, Lake Alfred pp. 2-7.
- Leite, R.P., and S.K. Mohan. 1990. Integrated management of the citrus bacterial canker disease caused by *Xanthomonas campestris* pv. *citri* in the State of Parana, Brazil. *Crop Protection* **9**: 3–7.
- Schaad, N.W. A.K. Vidaver, G.H. Lacy, K. Rudolph and J.B. Jones 2000. Evaluation of proposed amended names of several Pseudomonads and Xanthomonads and recommendations. *Phytopathology* **90**: 20-213.
- HMG/N, 2004/2005. Statistical information on Nepalese agriculture. *His Majesty's Government of Agriculture and Co-operative*. Agriculture-Business Promotion and Statistics Division, Singha Durbar, Kathmandu, Nepal pp. 1-47.
- Stall, R.E., J.W. Miller, G.M. Marco and B. Canteros 1980. Population dynamics of *Xanthomonas citri* causing canker of citrus in Argentina. *Proceeding of Fla. state horticulture society* **93**: 10-14.
- Stall, R.E., J.W. Miller, G.M. Marco and B. Canteros. 1982b. Timing of sprays to control canker of grapefruit in Argentina. *Proceedings of International society of citriculture* **1**: 414–417.
- Stall, R.L. 1993. Canker In Compendium of citrus diseases (Eds. J.D. Whiteside, M. Garnsey, L.W. Timmer). *The American Phytopathological society* pp 6-7.
- Vauterin, L., B. Hoste, K. Kersters and J. Swings 1995. Reclassification of *Xanthomonas*. *International Journal of Systematic Bacteriology* **45**: 472-489.
- Young, J.M., C.T. Bull, S.H. De Boer, G. Firrao, L. Gardan, G.E. Sandler, D.E. Stead and Y Takikawa 2001. Classification, nomenclature, and plant pathogenic bacteria. A clarification. *Phytopathology* **91**: 617– 620.

