Prevalence of Extended Spectrum Beta Lactamase Producing and Carbapenems Resistance Isolates from Hands of Health Care Workers in a Health Care Setting of Nepal

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ABSTRACT

Background: Hands of healthcare workers (HCWs) could be colonized by potential drug resistant bacteria like Extended Spectrum Beta Lactamase producers (ESBLs) and Carbapenems-resistant (CR) isolates and could become vectors of nosocomial pathogens in healthcare facilities that are associated with an increase of morbidity, mortality and healthcare costs. This study aimed to investigate the prevalence of ESBLs and CR isolates from hands of HCWs with their antibiotic susceptibility pattern. Materials and methods: This was a cross-sectional study that included a total of 150 hand swabs collected from March, 2018 to September, 2018 in Gandaki Medical College and Teaching Hospital. Isolation, identification and antimicrobial susceptibility tests were done using standard microbiological procedures. Results: Among the total isolates of 219 obtained from growth positive samples 92/219(42.01%) were Gram negative bacteria (GNB) and the most common were Klebsiella spp 32(34.78%) followed by Escherichia coli 17(18.48%), Pseudomonas aeruginosa12 (13.04%), Acenetobacter spp 11(11.96%), Proteus spp 9(9.78%), Citrobacter spp 7(7.61%) and Enterobacter spp 4(4.35%). The prevalence of ESBLs, CR and ESBLs with Co-resistant to Carbapenems were 19.56%,14.13% and 9.78%, respectively. The most effective drugs for isolates were Nitrofurantoin followed by Amikacin, Tetracycline and Gentamycin. Distribution pattern of the ESBLs and CR isolates among doctors, nurses, laboratory technicians, helpers and basic science faculties were not significant (p>0.05). Conclusions: This report revealed the emerging and moderately high prevalence of ESBLs, CR and ESBLs with Co-resistant to Carbapenems GNB with their antibiotic susceptibility patterns found on hands of HCWs in Nepal. Thus, this study could be helpful in developing proper guidelines on hand hygiene and implementation of infection control measures including contact precautions against the spread of infections by such pathogens in healthcare settings.

Key Words: Carbapenems-resistance, Extended Spectrum Beta Lactamase producers, Healthcare workers

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Submitted: April 13, 2020 **Accepted:** June 18, 2020

To cite: Sharma BK. Prevalence of Extended Spectrum Beta Lactamase Producing and Carbapenems Resistance Isolates from Hands of Health Care Workers in a Health Care Setting of Nepal. JGMC Nepal. 2020;13(1):23-31. **DOI:** 10.3126/jgmcn.v13i1.28461

INTRODUCTION

β-lactamases are a heterogeneous group of enzymes able to inactivate penicillins, cephalosporins and monobactams. These enzymes, frequently produced by aerobic and anaerobic Gram-negative bacteria, hydrolyze the β-lactam ring by irreversible hydroxylation, consequently inactivating the antibiotic. Enterobacteriaceae that produce extended spectrum β-lactamases carry plasmid-encoded enzymes that can efficiently hydrolyze and confer resistance to a variety of β-lactam antibiotics. ESBL-producing organisms are responsible for a significant proportion of infections in health care settings. Treatment of these infections can place an added constraint on already overburdened health systems in developing countries.

Infections with carbapenem-resistant *Enterobacteriaceae* (CRE) are increasingly being reported from patients in healthcare settings. They are associated with high patient



morbidity, attributable mortality and hospital costs. The management of infections has been complicated by the emergence of carbapenems resistance pathogens. Carbapenemases are β -lactamases that hydrolyse carbapenems, usually along with other β -lactams. The most frequently occurring species of *Enterobacteriaceae* which are found to be carbapenem-resistant and that produce carbapenemases, are *Klebsiella pneumoniae* and Escherichia coli.

Enterobacteriaceae can also produce β -lactamase enzymes such as AmpCs (chromosomal or acquired) and ESBLs that do not readily inactivate carbapenems on their own but can confer carbapenem resistance when combined with chromosomal porin mutations that prevent accumulation of β -lactam agents in the bacteria. Finally, the production of carbapenemase enzymes, typically found on mobile genetic elements that inactivate carbapenem and other

β-lactam antibiotics is increasingly common.^{8,9} These carbapenemase-producing CRE (CP CRE) frequently carry multiple resistance mechanisms, which can include redundant β-lactamases such as AmpCs and ESBLs and genes conferring resistance to other antimicrobial classes. While *Proteus, Providencia*, and *Morganella species* demonstrate intrinsically elevated MICs to Imipenem.¹⁰ β-lactams drugs are often the primary therapeutic option for serious infections, and carbapenems in particular areoften considered agents of last resort. Thus, the emergence and spread of ESBLs and carbapenem-resistant *Enterobacteriaceae* are significant clinical and public health concern.

Hand hygiene before patient contact is strongly recommended by the World Health Organization¹¹ and by Hand hygiene guidelines¹² as a measure to prevent crosstransmission of micro-organisms. Moreover, findings in different studies strongly recommend the implementation of multimodal campaigns to improve Hand hygiene compliance in HCWs and the use of sinkless alternatives such as alcohol based hand rub.^{12,13}

HCWs hands might be colonized with ESBLs and carbapenem-resistant Enterobacteriaceae isolates and contaminated hands of healthcare providers may cause threat to not only the life of hospital staffs, patients and visitors but also to the people in the community. Hand hygiene is one of the most important preventive interventions against the spread of infections in healthcare settings. There is a paucity of documented literature on investigation of ESBLs and Carbapenems resistance Gram negative organisms on hands of HCWs from Nepal. To the best of our knowledge this is the first report

from Nepal with an exclusive focus on investigating the current prevalence and of ESBL producing, Carbapenems resistance and ESBLs with Co-resistance to Carbapenems Gram negative organisms carriage among hands of HCWs from Nepal. Therefore, the study aimed to investigate the prevalence of ESBLs and CR isolates from hands of HCWs with their antibiotic susceptibility pattern.

MATERIALS AND METHODS

Study design and setting

This cross-sectional study was carried out at the Microbiology Laboratory from March, 2018 to September, 2018 after obtaining ethical clearance from Institutional Ethical Committee of Gandaki Medical College and Teaching Hospital. Verbal consent was taken from each participant and all samples were collected after he/she accepted and knew that they were participating in clinical study. The total of 150 hand swab samples were collected from the hands of Health care workers which include doctors, nurses, laboratory technicians and helpers working at various wards and departments of Gandaki Medical College and Teaching Hospital, Prithivichowk, Pokhara, Nepal.

Collection and Processing of Samples

HCWs working at various wards and departments and basic science faculties of Gandaki Medical College and Teaching Hospital were enrolled in this study who were apparently healthy and not taking any antibiotics two weeks prior to this study. The health care worker's hand swab samples were collected by means of sterile cotton swabs moistened in sterile saline water (0.85%). The sterilized cotton buds were rotated onto the overall surface area of palms of both hands and in between of the fingers too. The cotton bud swabs after swabbing the hands were kept in the sterile small tube containing Brain Heart Infusion (BHI) broth separately, labeled and was immediately transported to the microbiology laboratory of Gandaki Medical College and Teaching Hospital (GMC) for further processing.

All the swabs were cultured directly on MacConkey agar, Blood agar and Nutrient agar (Himedia). All cultured plates were incubated aerobically at 37°C for 24 hours. Bacterial isolates were identified using standard microbiological techniques. ¹⁴ Antimicrobial susceptibility testing of the isolates was performed by Kirby-Bauer disc diffusion technique and the interpretation was done according to Clinical and Laboratory Standards Institute (CLSI) guidelines (2017). ¹⁵

Screening and Confirmation of ESBL

The ESBL phenotype of identified colonies of Enterobacteriaceae were detected by confirmatory double disk diffusion test according to the CLSI guidelines (2017).¹⁵ For this, Cefotaxime (30 µg) or Ceftazidime (30 µg), discs were placed on Mueller-Hinton agar plates, 20 mm apart (centre to centre) from a Cefotaxime plus Clavulanic acid (30+10 mcg) and ceftazidime plus clavulanic acid (30+10 mcg). The zones of inhibition for the Cefotaxime and Ceftazidime alone was compared to that of combined disc of cefotaxime and Cefotaxime plus Clavulanic acid and Ceftazidime plus Clavulanic acid respectively. An increase of \geq 5mm in a zone of inhibition for either antimicrobial agent tested in combination with clavulanate vs zone of the inhibition of the agent when tested alone, was confirmed to be the ESBL producer. Plates were incubated overnight (18-20 h) at 35°C.

Carbapenem Susceptibility Testing

Susceptibility of all the isolates to carbapenems was tested in accordance with the current CLSI guidelines (CLSI 2012). Carbapenem antibiotics Meropenem and Imipenem were used. The discs were placed on the surface of inoculated Mueller Hinton Agar (MHA) (Hi media, India) plates using sterile forceps. The discs were placed about 25mm apart, and the plates were incubated for 24hours at 37°C after which zones of inhibition in diameter were recorded according to CDC guidelines (CDC 2009). It Isolates that showed a zone of inhibition <21mm in diameter for Meropenem or <23mm in diameter for Imipenem were considered as carbapenem resistant (CDC 2009).

Data Analysis

All the data were entered into a computer database using standard format, checked for errors and verified. Data maintained in the computer sheets were organized and analyzed by using GraphPad Prism software for Windows (version 8). A value of p≤0.05 was assumed wherever applicable and 95% confidence intervals along with the exact p-values were presented. Data were presented in appropriate table, figures by calculating percentage, rate etc. Appropriate statistics were applied wherever applicable.

RESULTS

The total of 150 hand swab samples were collected from the hands of health care workers which include doctors 33.33% (50/150), nurses 33.33% (50/150), helpers 13.33% (20/150), laboratory technicians 6.66% (10/150) working at various wards and departments of hospital and

13.33% (20/150) from hands of basic science faculties working at basic science blocks, Rithhepani Lekhnath Pokhara who does not have regular exposure to hospital environment of Gandaki Medical College and Teaching Hospital.

The frequency of bacterial growth positive among the cultivated hand swabs were 60% (90/150) and 40% (60/150) hand swab samples didn't show any growth. In most of the positive samples, they showed polymicrobial growth. Among the total isolates of 219 obtained from overall growth positive samples in this study 92/219 (42.01%) isolates were Gram negative bacteria and 127/219 (57.99%) were Gram positive bacteria. Bacterial isolates were identified using standard microbiological techniques.

Distribution Pattern of Gram-Negative Bacteria Isolated From Hand Swab Samples

The total numbers of Gram-negative bacteria isolated in this study was 92 isolates from 90 positive hand swab samples. The most common Gram-negative organism isolated in this study were *Klebsiella spp* 32 (34.78%) followed by *Escherichia coli* 17 (18.48%), *Pseudomonas aeruginosa* 12 (13.04%), *Acenetobacter spp* 11 (11.96%), *Proteus spp* 9 (9.78%), *Citrobacter spp* 7 (7.61%) and *Enterobacter spp* 4 (4.35%) (Table 1, figure 1).

Table 1. Distibution of Gram-negative bacteria isolated from various hand swab samples.

Organism Identified	Number	Frequency
Klebsiella spp	32	34.78%
Escherichia coli	17	18.48%
Pseudomonas aeruginosa	12	13.04%
Acenetobacter spp	11	11.96%
Proteus spp	9	9.78%
Citrobacter spp	7	7.61%
Enterobacter spp	4	4.35%
Total	92	100%

Distribution Pattern of ESBLs Isolated Gram Negative Bacteria

Out of total 92 Gram negative isolates, 18 were found to be ESBLs. The overall prevalence of ESBLs in this study was (18/92) 19.56%. Out of total isolates *Klebsiella spp* 6/92(6.52%); *Escherichia coli*, 4/92(4.35%); *Pseudomonasaeruginosa* 3/92 (3.26%); *Acenetobacter spp* 3/92 (3.26%) and *Proteus spp* 2/92 (2.17%) were found to be Extended B-Lactamase Producers. None of the isolated *Citrobacter spp* and *Enterobacter spp* was found to ESBLs producers (Table 2, figure 1).

Table 2. Distribution of ESBLs among the isolated Gram negative bacteria

Organism	Number of Isolates Tested	ESBL Producer N (%)		
Klebsiella spp	32	6(6.52%)		
Escherichia coli	17	4(4.35%)		
Pseudomonas aeruginosa	12	3(3.26%)		
Acenetobacter spp	11	3(3.26%)		
Proteus spp	9	2(2.17%)		
Citrobacter spp	7	-		
Enterobacter spp	4	-		
Total	92	18 (19.56%)		

Distribution Pattern of Resistance among the Isolated Gram Negative Bacteria to Carbapenems

Out of total 92 Gram negative isolates, 12 were found to be Carbapenems resistance (Resistant to both Meropenem and Imipenem). The overall prevalence of Carbapenem resistant to both Meropenem and Imipenem in this study was (13/92) 14.13%. Out of total isolates *Klebsiella spp* 4/92(4.35%); *Escherichia coli*, 3/92 (3.26%); *Pseudomonas aeruginosa* 3/92(3.26%); *Acenetobacter spp* 2/92 (2.17%) and *Proteus spp* 1/92 (1.09%) were found to be Carbapenems resistant. None of the isolated *Citrobacter spp* and *Enterobacter spp* was found to Carbapenems resistant (Table 3, figure 1).

Table 3. Distribution of Resistance among the isolated Gram negative bacteria to carbapenems

Organism	Number of Isolates Tested	No. resistant to Meropenem (Zones<21mm) N (%)	No. resistant to Imipenem (Zone<23mm) N (%)	No resistant to both N (%)	
Klebsiella spp	32	4 (4.35%)	6 (6.52%)	4 (4.35%)	
Escherichia coli	17	5 (5.43%)	3 (3.26%)	3 (3.26%)	
Pseudomonas aeruginosa	12	3 (3.26%)	3 (3.26%)	3 (3.26%)	
Acenetobacter spp	11	3 (3.26%)	2 (2.17%)	2 (2.17%)	
Proteus spp	9	1 (1.09%)	1 (1.09%)	1 (1.09%)	
Citrobacter spp	7	-	-	-	
Enterobacter spp	4	-	-	-	
Total	92	16 (17.39%)	15 (16.30%)	13 (14.13%)	

Distribution Pattern ESBLs Producers and Carbapenems Resistance Isolates

Out of total 92Gram negative isolates, 9 were found to be ESBLs Producers and Carbapenems resistance. The overall prevalence of ESBLs producers and Co-resistant to Carbapenems in this study was (9/92) 9.78%. Out of

total isolates *Klebsiella spp* 3/92 (3.26%); *Escherichia coli*, 2/92 (2.17%); *Pseudomonas aeruginosa* 2/92 (2.17%); Acenetobacter spp 1/92(1.09%) and *Proteus spp* 1/92 (1.09%) were found to be ESBLs producers and Coresistant to Carbapenems. None of the isolated *Citrobacter spp* and *Enterobacter spp* was found to ESBLs producers and Co-resistant to Carbapenems (Table 4).

Table 4. Distribution of ESBLs Producers and Carbapenems Resistance Isolates

Organism	Number of Isolates Tested	ESBL Producer + Carbapenems Resistance N (%)
Klebsiella spp	32	3 (3.26%)
Escherichia coli	17	2 (2.17%)
Pseudomonas aeruginosa	12	2 (2.17%)
Acenetobacter spp	11	1 (1.09%)
Proteus spp	9	1 (1.09%)
Citrobacter spp	7	-
Enterobacter spp	4	-
Total	92	9 (9.78%)

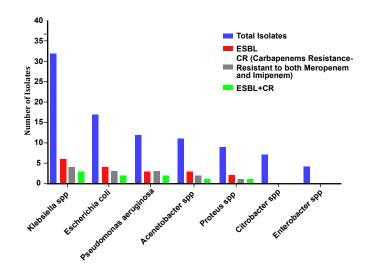


Figure 1: Distribution of ESBLs Producers, Carbapenems Resistance and ESBLs producers with Co-resistance to Carbapenems among the isolated Gram negative bacteria.

Antibiotic Susceptibility Test of the isolated Gramnegative bacteria

Various antibiotics were used for antibiotic susceptibility pattern determination using Kirby Bauer disc diffusion method. The most effective drug of choice for different isolates were Nitrofurantoin followed by Amikacin, Tetracycline and Gentamycin. All the isolates were 100% resistant to ampicillin and Amoxicillin+Clavulanate. Neither isolates of *Citrobacter spp* and *Enterobacter*

spp obtained in this study were resistant to Cefotaxime, Ceftazidime, Meropenem and Imipenem (Table 5).

Table 5. Antibiotic Susceptibility Test of the Isolated Gram-Negative Bacteria from Hand Swab Samples

Patho- gens	Kleb- siella spp	Esche- richia coli	Pseu- domo- nas aerugi- nosa	Acene- to- bacter spp	Prote- us spp	Citro- bacter spp	En- tero- bacter aero- gens
Total no. of isolates	32	17	12	11	9	7	4
	No. of iso	olates resi	stant to				
CTX	6 (18.75%)	4 (23.52%)	3 (25.0%)	3 (27.27%)	2 (22.22%)	0	0
CAZ	6 (18.75%)	4 (23.52%)	3 (25.0%)	3 (27.27%)	2 (22.22%)	0	0
MEM	4 (12.50%)	5 (29.41%)	3 (25.0%)	3 (27.27%)	1 (11.11%)	0	0
IPM	6 (18.75%)	3 (17.64%)	3 (25.0%)	2 (18.18%)	1 (11.11%)	0	0
AMP	0	0	0	0	0	0	0
AMC	0	0	0	0	0	0	0
CRO	26 (81.25%)	14 (82.35%)	10 (83.33%)	10 (90.90%)	8 (88.88)	6 (85.71%)	3 (75.0%)
NX	25 (78.12%)	12 (70.58%)	9 (75.0%)	9 (81.81%)	7 (77.77%)	5 (71.42%)	2 (50.0%)
GEN	17 (53.12%)	7 (41.17%)	4 (33.33%)	4 (36.36%)	4 (44.44%)	3 (42.85%)	2 (50.0%)
COT	24 (75.0%)	14 (82.35%)	8 (66.66%)	7 (63.63%)	7 (77.77%)	6 (85.71%)	3 (75.0%)
TE	10 (31.25%)	6 (35.29%)	5 (41.66%)	4 (36.36%)	3 (33.33%)	2 (28.57%)	2 (50.0%)
AK	8 (25.0%)	5 (29.41%)	3 (25.0%)	3 (27.27%)	2 (22.22%)	1 (14.28%)	1 (25.0%)
NIT	3 (9.37%)	3 (17.64%)	2 (16.66%)	2 (18.18%)	1 (9.09%)	1 (14.28%)	1 (25.0%)

CTX, Cefotaxime; CAZ, Ceftazidime; MEM, Meropenem; IPM, Imipenem; AMP, Ampicillin; AMC, Amoxicillin+Clavulanate; CRO, Ceftriaxone; NX, Norfloxacin; GEN, Gentamycin; COT, Cotrimoxzole; TE, Tetracycline; AK, Amikacin; NIT, Nitrofurantoin.

Distribution of Isolates of ESBLs Producers, Carbapenems Resistance and ESBLs producers with Co-resistance to Carbapenems in Relation to Samples Group

Among the total 92 Gram negative isolates obtained from different study groups total isolates were 18/92 (19.56%) from doctors, 29/92 (31.52%) from nurses, 19 (20.65%) from helpers, 15 (16.30%) from laboratory technicians and 11 (11.96%) from basic science faculties.Out of those tested 92 isolates 3 (3.26%) from doctors, 6 (6.52%) from nurses, 3 (3.26%) from helpers, 4(4.35%) from laboratory

technicians and 2 (2.17%) from basic science faculties that produced ESBLs, Among the 92 isolates tested, 2 (2.17%) from doctors, 5 (5.43%) from nurses, 2(2.17%) from helpers, 3 (3.26%) from laboratory technicians and 1 (1.09%) from basic science faculties were resistant to Carbapenems (Resistant to both Meropenem and Imipenem). Out of 92 isolates tested 2(2.17%) from doctors, 3(3.26%) from nurses, 1(1.09%) from helpers, 2(2.17%) from laboratory technicians and 1 (1.09%) from basic science faculties were ESBLs producers with Coresistance to Carbapenems. These data were not found in significantly different percentage from different study groups with p>0.05 (ESBLs, p=0.37; CR, p=0.40; ESBLs+CR, p=0.43) (Table 6).

Table 6. Distribution Pattern of Isolates of ESBLs Producers, Carbapenems Resistance and ESBLs producers with Co-resistance to Carbapenems in Relation to Sample Groups

	Doctors N (%)	Nurses N (%)	Helpers N (%)	Laboratory Techncians N (%)	Basic Science Faculties N (%)	Total N (%)	"p" value
		E	ESBLs Pro	ducers N (%)			
Total Isolates Tested	18	29	19	15	11	92	0.37
ESBL Positive (%)	3 (3.26)	6 (6.52)	3 (3.26)	4 (4.35)	2 (2.17)	18 (19.56)	0.37
Carbapene	ms Resista	nce (CR=	Resistanc	e to both Merc	penem and	Imipenem) N (%)
Total Isolates Tested	18	29	19	15	11	92	0.40
CR	2 (2.17)	5 (5.44)	2 (2.17)	3 (3.26)	1 (1.09)	13 (14.12)	0.40
ESBLs+CR (ESBLs Producers with Co-Resistance to Carbapenems) N (%)							
Total Isolates Tested	18	29	19	15	11	92	0.43
ESBL+CR	2 (2.17)	3 (3.26)	1 (1.09)	2 (2.17)	1 (1.09)	9 (9.78)	

DISCUSSION

Hand washing may not usually be performed often enough and many HCWs may not wash their hands properly in the course of their work throughout the day and contaminated hands of healthcare providers play a major role in spreading infections in healthcare settings. The emergence and spread of ESBLs and Carbapenems resistant bacteria through hands of HCWs is a public health threat as they usually associated with an increase of morbidity, mortality and healthcare costs. Hand hygiene is one of the most important preventive interventions against the spread of infections in healthcare settings.

In this study, the frequency of growth positive for bacteria among the cultivated hand swabs were 60% (90/150) and 40% (60/150) hand swab samples didn't show any growth. In most of the positive samples, they showed polymicrobial growth. The isolation rate of the pathogens was lower than previous study carried out by Kumari et al18 but comparable with the findings of Vedavati and Halesh¹⁹. Among the total isolates of 219 obtained from overall growth positive samples in this study 92/219 (42.01%) isolates were Gram negative bacteria and 127/219 (57.99%) were Gram positive bacteria. In this study, Gram positive bacteria were found to occur more than Gram negative bacteria. Most skin flora bacteria are Gram positive, which would account for their predominance on hand swab samples which was in accordance with the findings of Vedavati and Halesh¹⁹ who also demonstrated that the Gram positive bacteria predominate over Gram negative bacteria from the growth positive hand samples.

This study also highlighted the presence of potential pathogenic Gram-negative bacteria in hands of HCWs like Klebsiella spp 32(34.78%) followed by *Escherichia coli* 17(18.48%), *Pseudomonas aeruginosa* 12(13.04%), *Acenetobacter spp* 11(11.96%), *Proteus spp* 9(9.78%), *Citrobacter spp* 7(7.61%) and *Enterobacter spp* 4(4.35%). Similar types of pathogens were found on hands of health care workers in studies done by Vedabati and Halesh¹⁹ and Kumari et al¹⁸. The presence of these pathogenic bacteria on hands poses a potential risk to vulnerable, immunecompromised individuals. The fact that bacteria of the *Enterobacteriaceae* found on hands may indicate feacal contamination of the hands.

The overall prevalence of ESBLs in this study was (18/92) 19.56%. The frequency of ESBL producer in other studies was higher than our studies. 20,21 It may be due to steadily increasing the incidence of ESBL producing strains among the clinical isolates, also the prevalence of ESBLs among clinical isolates varies from country to country and from institution to institution.²² In the present study, Klebsiella spp was found to be the most common ESBL producer which was in concordance with the previous studies of Nepal and other countries.²³⁻²⁶ who had also reported that Klebsiella spp were the predominant ESBL producers among the clinical isolates. However, another study from Nepal has reported that Escherichia coli were the predominant ESBL producers among the clinical isolates.²⁰ In this study, none of the isolates of Citrobacter spp and Enterobacter spp were found to ESBLs positive which was in agreement with the findings of Lohani et al²⁰ who also reported none of the isolates of C. freundii were

ESBL positive in their study. It might be because of the less number of isolates.²⁷

The overall prevalence of Carbapenem resistant to both Meropenem and Imipenem in this study was (13/92) 14.13% which was lower as compared to prevalence of Carbapenem resistance from the family Enterobactericeae by Pokhrel et al²⁸. Among which most common bacteria was Klebsiella spp followed by Escherichia coli which was in agreement with the findings of previous studies done in Nepal by Pokhrel et al²⁸ and Bora et al²⁹ among the clinical isolates. None of the isolated Citrobacter spp and Enterobacter spp was found to be resistant to Carbapenems. In this study E. coli was more resistant to Meropenem followed by Klebsiella spp, Pseudomonas aeruginosa, Acenetobecter spp and Proteus spp. While, K. pneumoniae was more resistant to Imipenem followed by E. coli, Pseudomonas aeruginosa, Acenetobacter spp and *Proteus spp.* Poor hygienic practices by HCWs are some of the driving forces of ESBLs producers and Carbapenems resistance in this study. This is in accordance with the findings of Yusuf et al³⁰ who also demonstrated that E. coli was more resistant to Meropenem than to Imipenem while, in K. pneumoniae, the reverse was the case. The resistant genes in these pathogens could be easily transmitted not only to other HCWs, patients and visitors but also to the community if proper care is not taken.

The most effective drugs of choice for different isolates were Nitrofurantoin followed by Amikacin, Tetracycline and Gentamycin. A comparable finding was obtained by Chander and Shrestha²³ who also reported Nitrofurantoin and Aminoglycosides as optimal drugs of choice for isolated pathogens. Similarly Pokhrel et al²⁸ reported the increased rate of sensitivity of Amikacin in Carbapenem resistant enterobacteriaceae. All the isolates were 100% resistant to Ampicillin and Amoxicillin+Clavulanate. Neither isolates of Citrobacters pp and Enterobacter spp obtained in this study were resistant to Cefotaxime, Ceftazidime, Meropenem and Imipenem. The overall prevalence of ESBLs producers and Co-resistant to Carbapenems in this study was (9/92) 9.78%. Out of total isolates Klebsiellaspp 3/92 (3.26%); Escherichia coli, 2/92 (2.17%); Pseudomonas aeruginosa 2/92 (2.17%); Acenetobacter spp 1/92 (1.09%) and Proteus spp 1/92 (1.09%) were found to be ESBLs producers and Coresistant to Carbapenems. None of the isolated Citrobacter spp and Enterobacter spp was found to ESBLs producers and Co-resistant to Carbapenems. Though the prevalence is moderately high in this study, the simultaneous resistance to ESBL and Carbapenems by the isolates in

hands of HCWs make the whole scenario more alarming.

The health care workers harboring the ESBLs and Carbapenems resistant strains might be asymptomatic and could serve as carriers of these strains. Hand hygiene is one of the most important preventive interventions against the spread of infections by such pathogens not only to hospital settings but also to the community

Strengths and limitations

This study will be a useful reference for future studies, to explore and expand on the wider prevalence of ESBLs, Carbapenems resistant and ESBLs with Co-resistant to Carbapenems Gram negative organism found on hands of health care workers in Nepal. Since our study was based on phenotypic detection of ESBL production and Carbapenem resistance genotypic characterization is recommended in future studies.

CONCLUSION

The study identified a baseline of data on types of Gram negative bacterial isolates along with their antibiotic susceptibility patterns. This report revealed the emerging and moderately high prevalence of ESBLs, Carbapenems resistant and ESBLs with Co-resistant to Carbapenems Gram negative bacteria found on hands of health care workers in Nepal. Thus, this study could be helpful in developing proper guidelines on hand hygiene and implementation of infection control measures including contact precautions against the spread of infections by such pathogens in healthcare settings.

Conflict of Interest

The authors declare that they have no competing interests.

Acknowledgments

The author would like to express deep thanks and sincere appreciation to Dr. Keshab Sharma, Dr Madhu Timilsina and Ms. Sarita Adhikari for their help during sample collection. Authors would like to thank to the Dr. Sanjib Mani Regmi, laboratory technologist Mr. Bashanta Bhujel and all the faculty members of department of Microbiology, Gandaki Medical College and Teaching Hospital for their support.

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