

Research

Genetic diversity of *Ottelia acuminata* (Hydrocharitaceae) from the Eastern Himalayas, revealed by ISSR markers

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Abstract

Ottelia acuminata (Gagnep.) Dandy (Hydrocharitaceae), an endangered aquatic species, was investigated in the Eastern Himalayas, especially in Yunnan Province of Southwest China. The genetic diversity among seven populations was examined using inter-simple sequence repeat (ISSR) amplification markers. The field survey showed that 43.5% natural populations of *O. acuminata* have become extinct during the last 30 years. Among 13 remaining wild populations, eight (61.5%) are on the edge of extinction and only five (38.5%) were unaffected. For the study on seven populations based on ten primers, 147 clear and reproducible DNA fragments were generated, of which 144 (97.96%) were polymorphic. Within populations, however, the polymorphic bands (PPB) generated by ISSRs was 53 and occupied 36.05% in population B, and similarly within population J (51 and 34.69%, respectively). The results showed that genetic variation is much higher among populations of *O. acuminata* than within populations. Analyses of Nei's gene diversity, genetic distance and Shannon's index also agreed with these results. The average value of Nei's gene diversity (h) equaled 0.3710. The coefficient of genetic differentiation (G_{st}) equaled 0.5487, which means that 54.87% of the total molecular variance existed among populations. Such a high level of divergence present among populations may be caused by the complex topography and separated habitats which effectively restrict gene flow. Moreover, there is a lack of significant association between genetic and geographical distances ($r = 0.28889$, $P > 0.05$) in the populations of *O. acuminata*. Therefore, we proposed an appropriate strategy for conserving the genetic resources of *O. acuminata* in the Eastern Himalayas; namely, rescuing and conserving the core populations *in situ*, while selecting and preserving more populations with fewer individuals from each population *ex situ*.

Key-words: *Ottelia acuminata*, genetic diversity, conservation, Eastern Himalayas, Yunnan Plateau, China

Introduction

Ottelia (Hydrocharitaceae), with 21 species in the genus, is distributed in the warmer environment worldwide (13 species in Africa, seven in Asia and Oceania, and one in South America) (Cook *et al.* 1984). All the species in the genus are aquatic and are confined to freshwater. *Ottelia acuminata* is distributed in western China and occurs in Guizhou, Sichuan and Yunnan

provinces (Li 1981). It can be found in lakes, ponds and ditches. It can also tolerate slowly flowing water, and occurs in streams and rivers but absent in the fast flowing parts. Sometimes it can be grown in man-made waters, such as rice-fields, and pools for the cultivation of *Nelumbo* or *Trapa*. The altitude of its distribution areas varies from ca. 600 to 3,000 meters above sea level. Mostly it grows to a depth of 2.5 m under water surface. But in Luguhu Lake, where a variety occurs, *O. acuminata* var. *crispa*, it can grow up to 5 m deep (Li and Hsu 1979). The substrate is usually clay or

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sand, but in Luguhu Lake it grows in gravel. It is confined to calcareous waters with a pH of 7-8.4, and is absent from acidic or basic waters (Li 1987). The most frequent species associated with *O. acuminata* are *Potamogeton lucens* L., *Myriophyllum spicatum* L., *Potamogeton malaianus* Miq., *Potamogeton perfolianus* L., *Limnophila sessiliflora* Bl. and *Nymphaea tetragona* Georgi. In deep water the commonest associates are *Hydrilla verticillata* (L.f.) Royle, *Najas minor* (Pers.) All. and *Chara* sp. In most habitats *O. acuminata* is not present in large quantities (Li 1980). Usually it grows in groups of 2-3 individuals together. The leaves of such a group cover an area of ca. 2-4 m². The leaf laminas usually remain equally long on mature plants irrespective of the water depth, but the petioles vary enormously and may reach a length of 2-3 m (Cook and Urmi-Konig 1984). The laminas remain below the surface, while in shallow water the laminas may touch the surface of the water, but never floating. The flowers are borne above the water surface and the peduncles may reach the enormous length of 6 m. After fertilization the peduncles usually but not always become spiral and withdraw the developing fruits under water. Although the female spathes may contain up to 12 flowers, rarely more than 4 or 5 develop into ripe, seed-bearing fruits. The stem remains corm-like; there are no runners, stolons or specialized hibernacula, with the exception of a few individuals (in Changhu Lake and Xihu Lake) which bear bulbils in the male spathes (Li 1981; Li 1988; He 1991).

Ottelia acuminata is an ornamental, medicinal and tasty edible plant as well. The leaves, peduncles and spathes are gathered and sold in the markets to be eaten as vegetables by the local people. *O. acuminata* was the commonest aquatic plant on Yunnan Plateau 50 years ago (Ley et al. 1963). It is very sensitive to pollution, excessive collecting and the ravages of the imported grass carp (*Ctenopharyngodon idella* Val.). Since 1987, *Ottelia acuminata* has been recognized as an endangered species by the Chinese authorities (SEPA and IBCAS 1987). In recent years it has become extinct in some localities and very rare in others.

Thus, it is essential to protect this native and endangered species, and its genetic variation as well. Suitable molecular markers are necessary to reliably assess genetic diversity to investigate the population genetic structure of *O. acuminata* so that appropriate strategies can be adopted for its genetic improvement, restoration, conservation, and sustainable management.

The use of inter-simple sequence repeats (ISSRs) is a microsatellite-derived genetic fingerprinting method based on the amplification of DNA segments occurring in the genome in regions where a particular SSR (short sequence repeat) motif occurs on opposing strands within a short and amplifiable distance (Zietkiewicz et al. 1994). This method is similar to RAPDs, since both require no prior knowledge of the genome, cloning or specific primer design, yet it has higher reproducibility than RAPDs because of high annealing temperatures, and the cost of the analyses is lower than the cost of AFLPs (Zietkiewicz et al. 1994; Reddy et al. 2002). Therefore, ISSRs have been broadly and successfully used in studies on genetic diversity, phylogenetics, genetic mapping and evolutionary biology in a wide range of plant species (Ajibade et al. 2000; Chapman et al. 2000; Sankar and Moore 2001; Cheghamirza et al. 2004).

We have engaged in collection and genetic diversity analysis of *Ottelia acuminata* occurring in the Eastern Himalayas, especially on the Yunnan Plateau, since 2002. The present study included a detailed field survey of the population distribution of *O. acuminata* in Yunnan. And we tested the usability of ISSR primers to investigate the level and distribution of genetic diversity in seven natural populations of *O. acuminata*. Finally, we used 10 out of the 96 ISSR primers tested. The aim was to understand the changing distribution patterns, to reveal the genetic variation within and among populations of *O. acuminata* at the level of

Table 1. List of populations of *Ottelia acuminata* used in this study.

Population	Population localities	Water System	Longitude(E)/Latitude (N)
B	Heilongtan Spring, Baiyi village, Songming County, Yunnan	Jinsha River	102°52'/25°16'
N	Luguhu Lake, Ninglang County, Yunnan	Jinsha River	100°49'/27°42'
S	Changhu Lake, Shilin County, Yunnan	Zhu River	103°23'/24°42'
T	Beihai wetland, Tengchong County, Yunnan	Yiluowadi River	98°33'/25°07'
L	Lashihai Lake, Gucheng District, Yunnan	Jinsha River	100°09'/26°53'
G	Gemei River, Jianchuan County, Yunnan	Lancang River	100°11'/26°37'
J	Heilongtan Spring, Xinhua village, Heqing County, Yunnan	Jinsha River	99°55'/26°28'

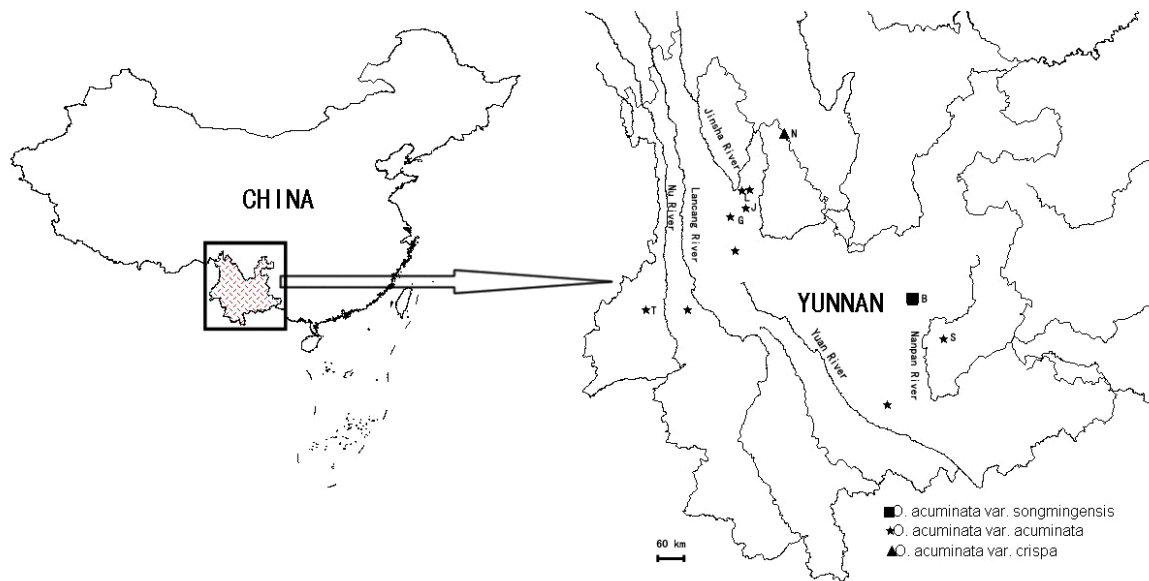


Figure 1. Distribution map of *Ottelia acuminata* populations.

DNA, and to find an effective method to conserve (both *in situ* and *ex situ*) and collect (sampling) this endangered species.

Materials and Methods

FIELD INVESTIGATION AND SAMPLING

From 2002 to 2005, we investigated the distribution pattern and extent of *O. acuminata* in Yunnan and found that the natural populations mainly remained in three regions: Lijiang, Dali and Kunming. According to the field survey, a total of 13 wild populations of *O. acuminata* have been approved to survive in the natural habitats (Figure 1). Seven natural populations of *O. acuminata* were sampled (Table 1). These populations were divided into four groups by water system: Jinsha River (i.e. the Yangtze) (B, N, L, J), Lancang River (the Mekong) (G), Zhu River (the Pear) (S) and Yiluowadi River (the Irrawaddy) (T). In each of the seven populations, 20 individual separated by a distance of more than five meters were collected and planted in a pool located at the Botanical Garden of the Kunming Institute of Botany.

TOTAL DNA EXTRACTION

Genomic DNA of the samples collected from seven populations was extracted using SDS improved protocol (Cheng *et al.* 2004). About 1.0 g dried leaf was ground in a

mortar with liquid nitrogen and incubated for 30 min at 65°C in 1.5 ml of SDS extraction buffer (100 mM Tris-HCl, pH 8.0, 500 mM NaCl, 50 mM EDTA, pH 8.0, 10 mM mercaptoethanol) and 600 µl 20% SDS. To the previous solution, 2 ml of 5 M potassium acetate (pH 7.5) was included and subsequently mixed. After 30 min incubation in an ice-water bath, the mixture was centrifuged at 11,400×g for 10 min at 4°C. The supernatant was mixed with an equal volume of chloroform–isoamyl alcohol (CI, 24:1) and centrifuged at 11,400 × g for 10 min at 4°C. The uppermost light solution was moved to a new tube and 0.6 volume cold isopropanol

Table 2. The nucleotide sequences of the ten selected primers, and the numbers of bands scored.

Primer	Primer sequence (5'-3')	Annealing temperature (°C)	Number of bands recorded
UBC816	(CA) ₈ T	52.7	14
UBC827	(AC) ₈ G	54.6	15
UBC846	(CA) ₈ RT	52.7	20
UBC849	(GT) ₈ YA	52.7	13
UBC850	(GT) ₈ YC	55.0	17
UBC855	(AC) ₈ YT	52.7	12
UBC856	(AC) ₈ YA	52.7	11
UBC857	(AC) ₈ YG	55.0	16
UBC873	(GACA) ₄	51.6	14
UBC889	DBD(AC) ₇	49.8	15

B = C/G/T; D = A/G/T; R = A/T; Y = C/G

was added. The tube was stirred carefully and put on the ice for 30 min for precipitation of the DNA. The DNA was recovered as a pellet by centrifugation at $11,400 \times g$ for 10 min at 4°C , washed twice with cold 70% ethanol. When the DNA pellet had air-dried at room temperature, it was dissolved in 200 μl TE (10 mM Tris-HCl, pH 8.0, 1 mM EDTA, pH 8.0) and 3 μl of 10 mg/ml of RNAase, and put in a water bath at 37°C for 1 h. Twenty microliters of 3M sodium acetate (pH 5.2) was added and then mixed. The DNA was again precipitated by adding 200 μl cold isopropanol. After centrifuging at $11,400 \times g$ for 10 min at 4°C , the pellet was washed with cold 70% ethanol, air-dried and resuspended in 100 μl TE buffer. The DNA samples isolated were diluted to 200 ng/ μl and stored at -20°C .

ISSR PCR AMPLIFICATION AND SELECTION OF OPTIMUM PRIMERS

A total of 96 ISSR primers obtained from Shengong Inc. were screened and 10 primers (Table 2) were selected for the ISSR analysis of *O. acuminata* based on the reproducible band. All amplifications were performed in an Eppendorf Mastercycler gradient PCR system (Eppendorf AG, 22331 Hamburg, Germany). The amplification reactions were performed in a volume of 25 μl containing 2 mM Mg^{2+} , 0.2 mM dNTP (Promega), 0.45 μM primer, 0.5 U Taq polymerase (Promega Co., USA), 2.5 μl of the $10\times$ reaction buffer and 20 ng of genomic DNA. PCR amplification was performed as follows: initial 5 min at 94°C , followed by 40 cycles of 45 s at 94°C , 45 s at $49-55^{\circ}\text{C}$, 90 s at 72°C , and a final 7 min extension at 72°C . The PCR products were separated on 1.5% agarose gels and stained by 0.1% ethidium bromide. The molecular weights were estimated using the E105-2 100 bp DNA Ladder Plus (Yunke Bio. Co. Ltd., China). The gel images were recorded and the band sizes were quantified using an E-box Fluorescence imaging system (Vilber Lourmat, France). The majority of the plant-primer combinations were run more than once to ensure reproducibility.

DATA ANALYSIS

ISSR amplified fragments, with the same mobility according to the molecular weight (bp), were scored manually for band presence (1) or absence (0). The following parameters were generated using the program POPGENE 1.32 (Yeh *et al.* 1997) to describe intra and inter-population genetic variation: Nei's gene diversity (h), Shannon's information index (I), the

observed number of alleles (na) and the effective number of alleles (ne) (Lewontin 1972; Nei 1973). Genetic divergence between populations was investigated using Nei's genetic distances (GD) and genetic identities (GID) (Nei 1978). Nei's genetic distances were calculated for all population pairs, using the NTSYS-pc v. 2.1 (Rohlf 1998) to construct a dendrogram using the unweighted pair group method (UPGMA). The genetic structure was further investigated using Nei's gene diversity statistics, including the total genetic diversity (Ht), genetic diversity within populations (Hs), and the relative magnitude of genetic differentiation among populations (Gst) (Nei 1973). An estimate of gene flow among populations (Nm) was computed using the formula of McDermott and McDonald $Nm = (1-Gst)/2Gst$ (McDermott and McDonald 1993). Using the NTSYS-pc v. 2.1 to make a Mantel test (Mantel 1967), it was used to test whether the matrix of genetic distances correlated with the matrix of geographical distances.

Results

During 2002–2005, we have made an extensive survey on *O. acuminata* in the Eastern Himalayas, especially on the Yunnan Plateau, including its current distribution, population characteristics and status of endangerment. The field survey indicated that there are 13 wild populations of *O. acuminata* remaining. These populations occupied an area ranged from 50 to 50,000 m^2 with an elevation between 1,515 and 2,730 m. Contrasting to 23 population numbers recorded during 1978–1982 (Li 1980; Li 1987), 10 (43.5%) natural populations of *O. acuminata* on Yunnan Plateau had disappeared in the past 30 years. In the existing 13 populations, 8 (61.5%) have been disturbed seriously and are on the verge of extinction, and only 5 (38.5%) grow normally.

A summary of the genetic data for between seven *O. acuminata* populations is given in Table 3. Among seven populations, 10 ISSR primers amplified 147 bands and 144 bands (PPB 97.96%) were found to be polymorphic. A total of 147 bands ranging from 200 to 1,800 bp were scored, corresponding to an average of 14.7 bands per primer. In individual populations, the percentages of polymorphic bands (PPB) ranged from 34.69% to 55.78%, with an average of 46.74% (Table 3). Nei's gene diversities (h) varied from 0.1072 to 0.2062, with an average of 0.1675, and Shannon's indices (I) ranged from 0.1647 to 0.3045, with an average of 0.2493.

Table 3. The genetic diversity of *Ottelia acuminata* and genetic structure of its populations.

Population	<i>na</i>	<i>ne</i>	<i>h</i>	<i>I</i>	<i>PPB</i>	<i>Ht</i>	<i>Hs</i>	<i>Gst</i>	<i>Nm</i>
B	1.3605	1.2203	0.1303	0.1943	36.05				
N	1.4694	1.2945	0.1714	0.2542	46.94				
S	1.5510	1.3593	0.2062	0.3045	55.10				
T	1.5306	1.3092	0.1802	0.2699	53.06				
L	1.5578	1.3560	0.2048	0.3033	55.78				
G	1.4558	1.2982	0.1721	0.2544	45.58				
J	1.3469	1.1737	0.1072	0.1647	34.69				
Mean	1.4674	1.2873	0.1675	0.2493	46.74				
Total	1.9796	1.6514	0.3710	0.5462	97.96	0.3710	0.1674	0.5487	0.4113

na = Observed number of alleles; *ne* = Effective number of alleles; *h* = Nei's (1973) gene diversity; *I* = Shannon's Information index; *PPB* = percentage of polymorphic bands; *Ht* = Total genetic diversity for the species; *Hs* = The mean heterozygosity within populations; *Gst* = Coefficient of gene differentiation; *Nm* = Estimate of gene flow.

Table 4. Nei's genetic identity (above diagonal) and genetic distance (below diagonal) among 7 natural *Ottelia acuminata* populations by ISSR markers.

Population	B	N	S	T	L	G	J
B	*	0.6889	0.6977	0.6621	0.6711	0.6434	0.6235
N	0.3726	*	0.7295	0.6502	0.7587	0.6988	0.6820
S	0.3600	0.3154	*	0.7868	0.8006	0.6882	0.7366
T	0.4123	0.4304	0.2398	*	0.7621	0.6980	0.6802
L	0.3988	0.2761	0.2224	0.2716	*	0.7680	0.8055
G	0.4409	0.3584	0.3737	0.3596	0.264	*	0.7997
J	0.4724	0.3827	0.3057	0.3854	0.2162	0.2236	*

Table 5. Geographic distance among 7 natural *Ottelia acuminata* populations.

Populations	B	N	S	T	L	G	J
B							
N	342.0						
S	84.9	423.8					
T	435.6	365.5	492.2				
L	329.4	114.0	410.8	249.8			
G	324.5	162.5	387.4	205.5	47.2		
J	309.0	135.9	391.3	232.8	28.8	27.5	

Moreover, the values of the parameters *h* and *I* showed a similar trend. The mean observed number of alleles (*na*) ranged from 1.3469 to 1.5578, while the mean effective number of alleles (*ne*) varied from 1.1737 to 1.3593. When calculated across populations, the *h* and *I* values equaled 0.3710 and 0.5462, respectively, and the *na* and *ne* values equaled 1.9796 and 1.6514, respectively. Among the seven populations investigated, population S exhibited the highest level of

variability while population J possessed the lowest value of variability.

The analysis of the population genetic structure revealed a considerable level of genetic differentiation among the seven populations of *O. acuminata* investigated. The total gene diversity (*Ht*) and gene diversity within populations (*Hs*) were 0.3710 and 0.1647, respectively. The coefficient of genetic differentiation (*Gst*), equaling 0.5487, indicated a high

degree of genetic differentiation among populations. The level of gene flow (Nm , the number of migrating individuals among populations per generation) was estimated to be only 0.4113.

The genetic distances, based on the allele frequencies of the ISSR markers, were calculated for each pair of populations to estimate the extent of their divergence (Table 4). The average genetic distance between populations was equal to 0.3372. The greatest genetic distance values were found between population B and other populations. This result revealed that population B, located geographically the furthest away from the other populations, is also genetically the most distant. However, Mantel's test showed that there was no significant correlation between genetic and geographical distances (Table 5) ($r = 0.28889$, $P > 0.05$) among the seven populations of *O. acuminata*.

A cluster analysis (UPGMA) was used to generate a dendrogram based on Nei's genetic distances of ISSR markers among seven populations (Figure 2). The dendrogram indicated that the population B of var. *songmingensis* and the population N of var. *crispa* was clearly separated from all other populations, while the five populations of var. *acuminata* clustered as a clade. The two populations (L, J) from Jinsha River clustered together before forming a cluster with any other population. In each cluster, individuals from the same population formed a distinct group. Overall, clusters were not related to the geographic distance between populations.

Discussion

ENDANGERED STATUS AND REASON OF *O. ACUMINATA*

According to the survey during 1978–1982 (Li 1980; Li 1987), 23 natural populations of *O. acuminata* had been recorded in Yunnan. For preserving and utilizing the plant genetic resources, we investigated again the numbers, localities, population and status of endangerment from 2002 to 2005. This work showed that 13 wild populations remained and 10 had vanished. In addition, 8 surviving populations were seriously disturbed and endangered; only 5 maintained normal growth. The study also indicates that the factors causing the species endangered are mainly: reclaiming from lake marshes, waters pollution, excessive fishes farming or collecting in lakes, water conservancy projects, lake drought and excessive vegetable collecting. Therefore, we should preserve this threatened species, based on a better understanding of genetic diversity among and within populations is essential for its conservation.

GENETIC DIVERSITY AMONG AND WITHIN POPULATIONS

Yunnan is located in the Eastern Himalayas. It is one of the regions with the most plentiful biodiversity in China and has a wide range of climatic and geographic conditions. According to our field investigation, the ecological environments and geographic distribution patterns of *O. acuminata* in Yunnan

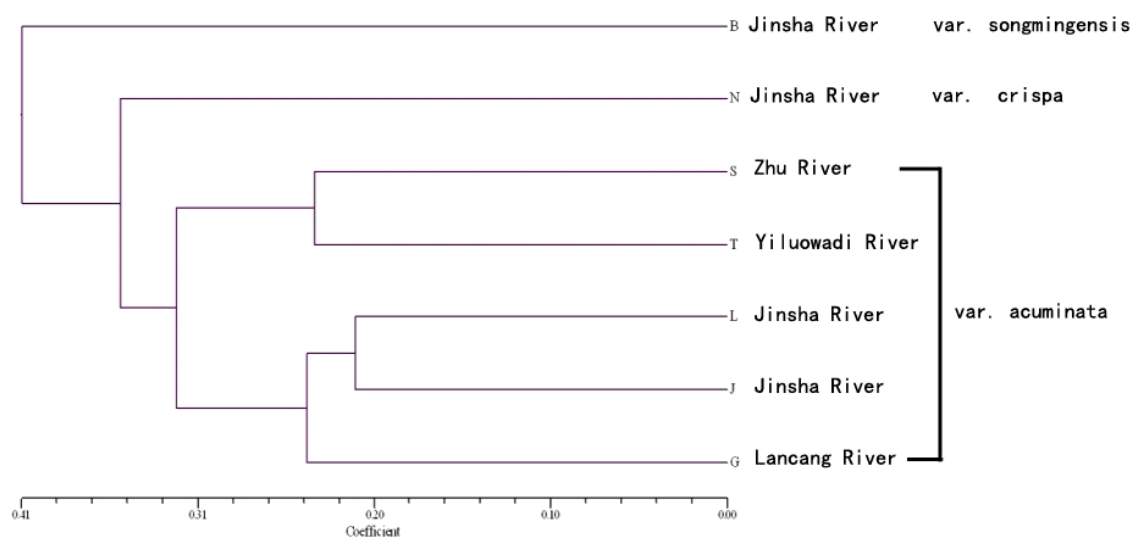


Figure 2. UPGMA dendrogram of populations of *Ottelia acuminata* based on Nei's genetic distance.

are diverse. Individuals among populations have different morphological characteristics. By using ISSR markers, this study showed that there is a high level of genetic diversity among populations of *O. acuminata* from Yunnan. This result agrees with previous studies based on allozyme analysis (He and Sun, 1992). Between seven populations, the percentages of polymorphic bands detected by ISSRs were 147 and 97.96%. And relatively low genetic diversity existed within populations where PPB values ranged from 34.69% to 55.78%, with an average of 46.74%, implying that a larger proportion of variation resided among populations. Similarly, Nei's gene diversity (0.37), unbiased measures of genetic distance and Shannon's information index (0.55) between populations were higher than that within populations. These analyses show that most of the genetic variations are partitioned between populations rather than within populations. The geographical and ecological differences in the distribution of genetic diversity of populations are extremely common. Many studies have clearly demonstrated that there is a clear association between population characteristics and the environments in which they occur (Aston and Bradshaw 1966; Al-Hiyaly *et al.* 1993). However, in this case, the evident genetic differentiation among populations of *O. acuminata*, genetic differentiation among populations does not appear to be correlated with geographic distance among the populations.

A high level of population differentiation may be explained by several factors, including the species' breeding system, genetic drift or genetic isolation of populations (Hogbin and Peakall 1999). When populations are small and isolated from one another, genetic drift influences genetic structure and increases differentiation among populations (Barrett and Kohn 1991; Ellstrand and Elam 1993). Low levels or absence of gene flow among populations is characteristic of many rare species (Slatkin 1985). Consequently, genetic differentiation among populations has been reported for a number of them (Sun and Wong 2001; Li *et al.* 2002). N_m , the effective gene flow per generation ($N_m = 0.4113$), for *O. acuminata* was lower than one successful migrant per generation, indicating limited gene flow among populations. The low rate of natural recruitment observed today, together with increased habitat fragmentation and large distances between populations is seriously contributing to the substantial decrease or even lack of gene flow at the population level.

Studies on habitat fragmentation indicate that this process can lead to population extinction and the loss of genetic variation, by not only minimizing suitable habitats, but also

increasing the mating opportunity between genetically closely related individuals. All of these factors could have contributed to a low level genetic diversity within populations of *O. acuminata* in the Eastern Himalayas.

Conclusion

A better understanding of the extent and patterns of genetic diversity of *O. acuminata* in the Eastern Himalayas, especially on the Yunnan Plateau, is essential for its conservation and sustainable uses. It will help us in determining what to conserve, and where and how to conserve this endangered species. The results from this work show that there is a high level of genetic diversity between populations but a low level genetic variation within populations. The loss of genetic diversity with extinction of populations is ever increasing.

We suggest that *in situ* conservation areas for five core *O. acuminata* populations growing normally should be established. A rational way for *ex situ* conservation is to collect and preserve the germplasm resources from eight populations disturbed seriously with relatively fewer individuals in each population.

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References

- Ajibade S.R., Weeden N.F. and Chite S.M. 2000. Inter simple sequence repeat analysis of genetic relationships in the genus *Vigna*. *Euphytica* 111: 47–55.
- Al-Hiyaly S.A.K., McNeilly T. and Bradshaw A.D. 1993. The effect of zinc contamination from electricity pylons: genetic constraints on selection for zinc tolerance. *Heredity* 70: 22–32.

- Aston D.L. and Bradshaw A.D. 1966. Evolution in closely adjacent populations. Part II. *Agrostis stolonifera* in maritime habitats. *Heredity* 37: 9–25.
- Barrett S.C.H. and Kohn J.R. 1991. Genetics and evolutionary consequences of small population size in plants: implications for conservation. In: *Genetics and Conservation of Rare Plants* (D.A. Falk and K.E. Holsinger, eds.), pp. 3–30. Oxford University Press, NY, USA.
- Chapman H.M., Parh D. and Oraguzie N. 2000. Genetic structure and colonizing success of a clonal, weedy species, *Pilosella officinarum* (Asteraceae). *Heredity* 84: 401–409.
- Cheghamirza K., Koveza O.V., Konovalov F.A. and Gostimskii S.A. 2004. Identification and mapping of chi115 gene and DNA markers linked to it in pea (*Pisum sativum* L.). *Genetika* 40: 909–915.
- Cheng J.W., Zai Q.C., Xing Q.H., Shou H.Y., Kai M.C. and Chong R.S. 2004. Genetic diversity among and within populations of *Oryza granulata* from Yunnan of China revealed by RAPD and ISSR markers: implications for conservation of the endangered species. *Plant Science* 167: 35–42.
- Cook C.D.K. and Urmi-Konig K. 1984. A revision of the genus *Ottelia* (Hydrocharitaceae): 2. the species of Eurasia Australasia and America. *Aquatic Botany* 20: 131–178.
- Cook C.D.K., Symoens J.J. and Urmi-Konig K. 1984. A revision of the genus *Ottelia* (Hydrocharitaceae): 1. generic considerations. *Aquatic Botany* 18: 263–274.
- Ellstrand N.C. and Elam D.R. 1993. Population genetics consequences of small population size: implications for plant conservation. *Annual Review of Ecology Evolution and Systematics* 24: 217–243.
- He J.B. 1991. *Systematic Botanical and Biosystematic Studies on Ottelia in China*. Wuhan University Press, Wuhan.
- He J.B. and Sun X.Z. 1992. Electrophoretic banding patterns of three isoenzymes of Chinese *Ottelia* and their systematic significance. *Journal of Wuhan Botanical Research* 10: 35–42.
- Hogbin P.M. and Peakall R. 1999. Evaluation of the conservation of genetic research to the management of endangered plant *Zieria prostrata*. *Conservation Biology* 13: 514–522.
- Lewontin R.C. 1972. The apportionment of human diversity. *Evolution Biology* 6: 381–398.
- Ley S.H., Yu M.K., Li K.C., Tseng C.M., Chen C.Y., Kao P.Y. and Huang F.C. 1963. Limnological survey of the lakes of Yunnan plateau. *Oceanologia et Limnologia Sinica* 5: 87–114.
- Li H. 1980. A study on the lake vegetation of Yunnan Plateau. *Acta Botanica Yunnanica* 2: 113–141.
- Li H. 1981. Classification, distribution and phylogeny of the genus *Ottelia*. *Acta Phytotaxonomica Sinica* 19: 29–42.
- Li H. 1987. The lake vegetation of Hengduan Mountains. *Acta Botanica Yunnanica* 9: 257–270.
- Li H. 1988. The aquatic vegetation and flora in Changhu Lake. *Journal of Yunnan University* Suppl. 10: 119–123.
- Li H., Hsu T.Z. 1979. The geobotanical expedition on Lake Luguhu. *Acta Botanica Yunnanica* 1: 125–137.
- Li Q.M., Xu Z.F. and He T.H. 2002. Ex situ genetic conservation of endangered *Vatica guangxiensis* (Dipterocarpaceae) in China. *Biological Conservation* 106: 151–156.
- Mantel N. 1967. The detection of disease clustering and a generalized regression approach. *Cancer Research* 27: 209–212.
- McDermott J.M. and McDonald B.A. 1993. Gene flow in plant pathosystems. *Annual Review of Phytopathology* 31: 353–373.
- Nei M. 1973. Analysis of gene diversity in subdivided populations. *Proceedings of the National Academy of Sciences of the United States of America* 70: 3321–3323.
- Nei M. 1978. Estimation of average heterozygosity and genetic distance from a small number of individuals. *Genetics* 89: 583–590.
- Reddy M.P., Sarla N. and Siddiq E.A. 2002. Inter simple sequence repeat (ISSR) polymorphism and its application in plant breeding. *Euphytica* 128: 9–17.
- Rohlf F.J. 1998. *NTSYS-Numerical Taxonomy and Multivariate Analysis System*. Exeter Publ., NY, USA.
- Sankar A.A. and Moore G.A. 2001. Evaluation of inter-simple sequence repeat analysis for mapping in *Citrus* and extension of the genetic linkage map. *Theoretical and Application Genetics* 102: 206–214.
- SEPA (State Environment Protection Administration), IBCAS (Institute of Botany, Chinese Academy of Sciences) 1987. List of Rare and Endangered Plants of China (vol. I). Science Press, Beijing.
- Slatkin M. 1985. Gene flow in natural populations. *Annual Review of Ecology Evolution and Systematics* 16: 393–430.
- Sun M. and Wong K.C. 2001. Genetic structure of three orchid species with contrasting breeding systems using RAPD and allozyme markers. *American Journal of Botany* 88: 2180–2189.
- Yeh F.C., Yang R.C., Boyle T.B.J., Ye Z.H. and Mao J.X. 1997. *POPGENE, the User-Friendly Shareware for Population Genetic Analysis*. Molecular Biology and Biotechnology Centre, University of Alberta, Edmonton, Canada.
- Zietkiewicz E., Rafalski A. and Labuda D. 1994. Genome fingerprinting by simple sequence repeat (SSR)-anchored polymerase chain reaction amplification. *Genomics* 20: 176–183.